

## Detection of *Cronobacter sakazakii* (*Enterobacter sakazakii*) in powdered food infants (PIF) and raw milk in Iraq

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### Abstract:

This study was conducted to detect *C.sakazakii* PIF and raw milk. Two hundred samples of PIF were taken from the infected hospital infants who used this type of milk and from the local markets in addition to 16 sample of raw milk were collected. The study is the first to report the isolation of *C. sakazakii* and *Enterobacter* spp. from raw milk in Iraq. The distribution of *C.sakazakii* and *Enterobacter* spp. among the presumptive isolates using Vitek-GN2 system gave 1/16(6.25%) isolates of *C.sakazakii* and 4/16 (25%) isolates of *Enterobacter* spp. *Enterobacter* spp. isolates include (*E.cloacae* ssp. *cloacae* and *E.cloacae* ssp. *dissolvens*, *E.hormaechei*, and *E.ludwigii*) that isolate from raw milk Differences in between percentages of each isolate persence were non-significant (P<0.05). The results of antibioticsusceptibility were determined using Vitek-2GN system; *.sakazakii* isolates showed 100% resistance to cefazolin and cefoxitin, but were highly sensitive to many antibiotics includes (Imipenem, Meropenem, Amikacin, Gentamicin, Tobramycin, Ciprofloxacin, Levofloxacin, Nitrofurantion, Trimethoprim \sulfamethoxazole, Ampicillin, Ampicillin\ sulbactam, Piperacillin \Tazobactam, Ceftazidime, Ceftriaxone, Cefepime Azetreonam and augmentin ). The present study did not determine *C.sakazakii* in all the samples of PIF that is available in the local markets.

**Key words:** *Cronobacter sakazakii*; powdered infant formula (PIF), milk, *Enterobacter* spp. .

### Introduction:

*Cronobacter sakazakii* (*C.sakazakii*) previously known as *Enterobacter sakazakii* (*E.sakazakii*) is a motile , Gram-negative , non sporing yellow pigmented rod , which belongs to lethal *Enterobacteriaceae* family ; It is an opportunistic human and food- born pathogen [1,2,3]. The organism can be found in broad range of foods including powdered infant formula (PIF) , cheese, meat, vegetables, grain, herbs , spices , tomato , water and households [2,4,5] . Also *C. sakazakii* is ubiquitously found in air, soil, floor drain and dry product processing environment. Due to its virulence *C. sakazakii* cause life threatening infection such as septicemia , and

necrotizing enterocolitis (NEC), bacteremia and meningitis in infants [1,6] .The mortality rates of 33 - 80% were reported among infected patients [7]. NEC is the most common gastrointestinal surgical emergency in neonatal populations, which results in a mortality rate of 40–100% in the most severely affected patients [8].Neonatal infections have been reported to be one rise via contact with *C.sakazakii* in the birth canal or through post-birth environmental sources [9]. The organism was resistant to multiple antibiotics and required prolonged treatment with broad spectrum antibiotics [10,11,12]. In recent years, the International Commission on

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Microbiological Specifications for Foods has ranked *C.sakazakii* a “severe hazard for restricted populations.” Because of its resistance to certain antibiotics [13]. In Iraq, *C.sakazakii* was isolated (5.71%) from 2 commercial samples of PIF from 35 total samples and this isolate produce heat-labile enterotoxin in mouse [14]. Both the source of *Cronobacter* and vehicle of transmission are not always clear. However, powdered infant formula (PIF) has been epidemiologically linked to the cases of infants infection [15], This study has focused on the surveillance of PIF products for the presence of *Cronobacter* compared to raw milk. The aims of the study are detection of *C.sakazakii* from PIF and raw milk from local Iraqi markets and food sample (PIF) from Iraqi infant's hospital patients.

## Materials and Methods:

### Samples and Isolation

A total of 216 food samples were collected at the following:-

**A.** Raw milk (16) samples were purchased from local markets, transported in a cool box and transport to the laboratory and testing it on the same day.

**B.** One hundred samples of PIF were collected from infected hospital infants patients (meningitis and NEC).

**C.** One hundred samples of PIF were purchased from the local markets. The samples were taken after recording the labeling information (name of company, commercial name of product, contents, origin, date of expired and production, batch number); then kept at room temperature and in dry condition during the study.

The samples were tested for the presence of *Cronobacter* spp. as

described by modified Chap *et al.*, [16]. Briefly, 25 g. of food samples (PIF, raw milk) were added to 225 ml of peptone water and then incubated at 37°C for 18-24 hrs. A 10 ml aliquot was then incubated in 90 ml Enterbacter enrichment broth (EEb) at 37°C for 18-24 hrs. From each enriched sample, 0.1 ml was streaked or spread onto Hicrome *Enterobacter sakazakii* Agar (HESA) .Up to five presumptive *C. sakazakii* colonies that exhibited during culture on HESA were selected for culturing on Trypton soy agar (TSA) at 25°C for 48-72 hrs. For identification use biochemical test, API-20E and Vitek-2 GN system were used (Biomerieux / France).

### Antimicrobial susceptibility test.

All the confirmed isolates were tested to antibiotic resistance using Vitek-2 GN system according to production company in AL.Mahmudia Hospital .The tested antibiotics (n=17) includes (Ampicillin, Ampicillin/sulbactam, Piperacillin \Tazobactam, Cefazolin, Cefoxitin, Imipenem, Meropenem, Amikacin, Gentamicin, Tobramycin, Ciprofloxacin, Ceftazidime, Ceftriaxone, Cefepime, Levofloxacin , Nitrofurantion, and Trimethoprim \sulfamethoxazole).

### Results and Discussion:

Table (1-1) shows 5 isolates (2.31%) which appears blue-black (n=2) to blue-gray (n=3) colonies on HESA , raised colonies 1-2 mm diameter , with and without halos after 18-24 hr. at 37°C and microscopic examination appears gram negative, straight rods, appeared as single or double chains and motile were considered presumptive *Cronobacter* spp. or presumptive *C.sakazakii* .

**Table 1-1: Percentages of presumptive *Cronobacter* spp. isolates and its characteristics on culture media according to the type of sample**

Samples (No.)		Characteristics on HESA (No. of presumptive <i>Cronobacter</i> spp. Isolates)	% of total samples	Characteristics on TSA (No. of presumptive <i>Cronobacter</i> spp. Isolates)	% of total isolate
Food samples (216)	Raw milk (16)	Blue-black ( 2 ) blue-gray ( 3 )	2.31	Yellow pigment ( 1 )	100
	PIF (200)	0	0.00	0	0.00
Total		5	2.31	1	100

HESA was developed for isolating presumptive colonies of *C. sakazakii*, this media contains two chromogenic substrates (5-bromo-4-chloro-3-indoxyl- $\alpha$ -D-glucopyranoside and 5-bromo-4-chloro-3-indoxyl- $\beta$ -D-cellobioside), three sugars (sorbitol, D-arabitol, and adonitol), a pH indicator, and inhibitors (bile salts, vancomycin, and cefsulodin), which all contribute to its selectivity and differential properties [17].

#### Biochemical tests

All the tested isolates (n=5) that gave yellow pigmented on TAS give the same result to the biochemical tests. The urease, oxidase, coagulase, indol and H<sub>2</sub>S production gives negative result, while catalase, citrate and motility test gives positive result. (Table 1-2). These results showed conformity with the *Cronobacter* spp. or *C.sakazakii* [3,18,19].

**Table 1-2: Biochemical tests of the presumptive *Cronobacter* spp. isolates**

Test	Results
Urease, Oxidase, Coagulase, Indol, H <sub>2</sub> S production	Negative
Catalase, Citrate, Motility	Positive

#### API 20 E test

The tested isolates (n=5) that gave positive results in the biochemical tests appeared different results in API 20E test. One isolate showed conformity with the *C.sakazakii*, while 4 isolates showed conformity with the *Enterobacter cloacae*. API 20E for

*C.sakazakii* revealed that ADH, ODC and CIT were positive, ONPG, H<sub>2</sub>S, LDC, URE, TDA and IND were negative. VP and GEL were also positive and all sugars GLU, MAN, RHA, SAC MEL, AMY and ARA were positive except SOR which was negative, While the results of API 20E for *Enterobacter cloacae* revealed that ADH, ODC and CIT were positive, ONPG, H<sub>2</sub>S, LDC, URE, TDA and IND were negative. VP and GEL were also positive and all sugars GLU, MAN, RHA, SAC, SOR, MEL, AMY and ARA were positive except INO which was negative.

The API 20E biochemical kit has been reported not to be a reliable tool for the confirmation of the identity of *C.sakazakii* [3,20]. Reported that the misidentification of strains by the API 20E biochemical kit was due to its limited biochemical gallery. The latter kit consisted of a wide variety of fermentable carbon sources and was able to correctly identify strain *E. cloacae* rather than as *C. sakazakii*. The correct identification of *C. sakazakii* based on biochemical profile kits has been reported to be problematic [20,21]. The fact that *C.sakazakii* confirmation should be based on more than one confirmation system. Both the API 20E and Biolog Microlog 34.20 systems should be used for confirmation of *C.sakazakii* isolates [17].

#### Vitek -2 system

Five presumptive isolates (on HESA) were tested by Vitek GN2 system. One

isolate that showed conformity with the *C.sakazakii* in API 20E test showed excellent identification confidence with the *C.sakazakii* organism in Vitek-GN2 with the identification values or probability 99%, while 4 isolate showed excellent identification confidence with the *Enterobacter* spp. that included *E.cloacae* spp. *dissolvens*, *E.cloacae* spp. *cloacae*, *E.hormaechei* and *E.ludwigii* with identification values (94 –99)% . Kim *et al.* [12] identified and conformed 4 isolates of *C.sakazakii* from clinical samples, three of the Vitek GNI biochemical profiles of isolates identified as *C. sakazakii* had identification values of 96 - 99% and that of one strain was 82%. Oonaka *et al.* [22] identified 52 strains of *Enterobacteriaceae* isolated from PIF using Vitek GN2 compact system (Biomerieux) with the identification values of 80 - 99%. From this, the results indicated that Vitek-2 system has the ability to distinguish between *Enterobacter* spp. (*E.cloacae* ssp. *cloacae* and *E.cloacae* ssp. *dissolvens*, *Enterobacter* spp, *E.hormaechei*, *E.kobei*, *E.ludwigii*) rather than as *C. sakazakii* and *E. cloacae* only , while API-20E system does not have the ability to distinguish between *E.cloacae* and other *Enterobacter* spp. Because of the phenotypic differences among *Cronobacter* spp., it has been increasingly difficult to confirm the identity of isolates using only one method or one set of *Cronobacter* spp.-specific PCR primers. [3,23] There are also problems with specificity, especially when discriminating between *Cronobacter* spp. and other *Enterobacter* species [24]. Therefore, a combination of confirmation methods might be necessary to completely eliminate false positives and false negatives. The results of this study on PIF differed from the study of Muytjens *et al.* [25] that examined 141

different powdered formulas from 35 countries and isolated *C. sakazakii* at levels ranging from 0.36 to 66 cfu per 100 g from 20 formula samples from 13 countries. Over a 24-month surveillance, three *Cronobacter* strains were isolated from 77 powder infant formulas (3.90%) and no *Cronobacter* was detected in liquid milk. [26]

As shown in Table (1-1) from 5 presumptive isolates (on HESA) only 1/16 (6.25) from raw milk. Steigerwalt *et al.*[27] have observed that the presence or absence of yellow pigmentation can distinguish between two strains of *Enterobacteriaceae* family (*Enterobacter cloacae* and *C.sakazakii*) when these strains are cultured on TAS. Although Farmer *et al.* [28] reported that not all strains of *C. sakazakii* are yellow-pigmented.

Eventhough *C. sakazakii* has been determined as a dominant contaminant flora in PIF by several researchers up to now, and in Iraq Shareef *et al.*[14] was isolated *C.sakazakii* 2/35(5.71% ) from PIF, one of them from sample opened at experimental time and the second from sample opened before 3 days of experimental time by using Blood agar and MacConkey agar with the traditional biochemical test . In the present study *C.sakazakii* was not determined in all samples (n= 200). Table (1-1) ,This result agrees with that mentioned by Block *et al.*[29], who stated that the organism was not isolated from PIF, but it was recovered from prepared formula and from a kitchen blender. The survey conducted by the South Australian Government did not find any pathogens in the 20 samples tested [30]. Also it coincides with what Sani and Yen Yi. [31] who reported that *C. sakazakii* was not detected in any of the PIF tested (30 samples) from 8 manufacturers obtained from hypermarkets and a private hospital in Malaysia. And coincide with Güner *et*

*al.*[32]who did not found *C. sakazakii* in all samples( 132 cartons) of PIF from 3 different brands retailed in Turkey . Joosten and Iversen [33]have detected 1.9% (n=104) *C. sakazakii* positive samples.

While PIF may not be commercially sterile, its production is undertaken using rigorous hygienic precautions coupled with monitoring of the process environment and finished product by the manufacturer. These activities assist in reducing the microbial load of

PIF [34]. The Codex Alimentarius Commission criteria require that no *C. sakazakii* should be present in 10 g of PIF after primary packaging until the opening of the can for consumption [35] A coordinated survey for *Cronobacter* and related organisms in PIF formula, follow up formula and infant foods was undertaken by 8 laboratories in 7 countries, *C. sakazakii* was isolated from 3/91(3%) follow up formulas [16].

**Table 1-3: Distribution of *Enterobacter* spp among the isolates according to type of samples using Vitek-2 system.**

Type of samples (No. of isolates)	<i>E.cloacae</i> sp. <i>Cloacae</i>	<i>E.cloacae</i> ssp. <i>dissolvens</i>	<i>E.hormaechei</i>	<i>E. ludwigii</i>	Chi-square value
Raw milk (4)	1(25)	1(25)	1(25)	1(25)	0.043 NS
Total (4)	1(25)	1(25)	1(25)	1(25)	

NS: Not-significant.

As shown in table 1-3 that indicates the presence of the common isolate *Enterobacter* spp. (*E.cloacae* ssp.*cloacae*, *E.cloacae* ssp. *dissolvens* *E.hormaechei*, *E.ludwigii*) from raw milk. This result agrees with Campos et al. [36] who has been shown to be of clinical significance by the report of several outbreaks of sepsis in neonatal intensive care units in Brazil and the USA that causes by *Enterobacter* spp. Differences between percentages of each isolate were non-significant(P<0.05).Species of the *Enterobacter cloacae* complex are widely encountered in nature, but they can act as pathogens. The biochemical and molecular studies on *E. cloacae* has shown genomic heterogeneity, comprising six species: *Enterobacter cloacae*, *Enterobacter asburiae*, *Enterobacter hormaechei*, *Enterobacter kobei*, *Enterobacter ludwigii* and *Enterobacter nimipressuralis*, *E. cloacae* and *E. hormaechei* are the most frequently

isolated in human clinical specimens [37].

### Antimicrobial susceptibility test

The results of antibiotic susceptibility have been that done using Vitek-2 system *C.sakazakii* isolate was 100% resistant to cefazolin and cefoxitin.*C.sakazakii* isolates are high sensitive to many antibiotics includes (Imipenem, Meropenem, Amikacin, Gentamicin, Tobramycin, Ciprofloxacin, Levofloxacin, Nitrofurantion, Trimethoprim \sulfamethoxazole, Ampicillin, Ampicillin \sulbactam, Piperacillin \Tazobactam, Ceftazidime, Ceftriaxone, Cefepime Azetreonam and augmentin). The results in the present study coincide with previous studies. Stock and Wiedeman [12] ; Drudy et al. [38] that has found *C.sakazakii* isolates locally shown 100% sensitive to ampicillin and gentamycin or ampicillin and chloramphenicol. Shadlia-Matug et al.[39] that have found *C.sakazakii*

resistance to cephalothin and sensitive to gentamicin, but not agreement with them that *C.sakazakii* resistance to ampicillin, penicillin. The present results also coincide with Stock and Wiedemann [12] found isolates were susceptible to ampicillin, compound sulphonamides, furazolidone, gentamicin, spectinomycin and streptomycin. In the present study *C.sakazakii* isolates have shown high sensitive to ampicillin, gentamicin this result coincides with pervious study [40,41] .And with Lai, [7] that found *C.sakazakii* were susceptible to Trimethoprim \sulfamethoxazole. The results of this study to not have agreement with Oonaka *et al.* [22] that found *C. sakazakii* were resistant to ampicillin and lincomycin, but they have agreement with them when found sensitive to gentamicin and cephalosporins. Also not agreement with Pitout *et al.*[42]; Girlich *et al.* [10] that they found *C.sakazakii* resistance to ampicillin .And with Kim *et al.*, [11] that tested 113 *C. sakazakii* isolates for their antibiotic resistance and 31.8% were resistant to ampicillin and all of the isolates were resistant to at least one antibiotic. The results of the present study coincide with pervious study by Al-Nabulsi *et al.*, [43] illustrated the *C. sakazakii* strains they tested were sensitive to gentamycin, kanamycin, ciprofloxacin and amoxicillin.

### Reference:

1. Arsalan, A.; Anwar, Z.; Ahmed, I.; Shad, Z.; and Ahmed, S. 2013.*Cronobacter sakazakii* an emerging contaminant in pedidtric infant milk formula.Int. Res. J.Pharm,4: 17-20.
2. Fiore, A; Casale, M; and Aureli, P.2008. *Enterobacter sakazakii*: epidemiology, clinical presentation, prevention and control. Ann Ist Super Sanità, 44(3): 275-280.
3. Iversen, C.; Lehner, A.; Mullane, N.; Bidlas, E.; Cleenwerck, I.; and Marugg, J.2007. The taxonomy of *Enterobacter sakazakii*: proposal of a new genus *Cronobacter* gen. nov. and descriptions of *Cronobacter sakazakii* comb. nov. *Cronobacter sakazakii* subsp. *sakazakii*, comb.nov., *Cronobacter sakazakii* subsp. *malonaticus* subsp. nov., *Cronobacter turicensis* sp. nov., *Cronobacter muytjensii* sp. nov., *Cronobacter dublinensis* sp. nov., *Cronobacter* genom species 1. BMC Evol. Biol., 64 (7): 1471-2148.
4. Beuchat, L.R.; Kim, H.; Gurtler, J.B.; Lin, L.C.; Ryu, J.H.; and Richards, G.M.2009. *Cronobacter sakazakii* in foods and factors affecting its survival, growth, and inactivation. Intl J Food Microbiol., 136 (2): 204-13.
5. Kandhai, M.C.; Reij, M.W.; Gorris, L.G.; Guillaume-Gentil, O.; and van Schothorst, M.2004.Occurrence of *Enterobacter sakazakii* in food production environments and households. Lancet., 363: 39–40.
6. Hunter, C.J.;and Bean, J. F. 2013. *Cronobacter*: an emerging opportunistic pathogen associated with neonatal meningitis, sepsis and necrotizing enterocolitis. J. of Perinat., V: 33(8) : 581-585.
7. Lai, K.K.2001. *Enterobacter sakazakii* infections among neonates, infants, children, and adults. Case reports and a review of the literature. Medicine (Baltimore), 80(2):113-22.
8. Stoll, B.J.1994. Epidemiology of necrotizing enterocolitis. Clin. Perinatol. 21:205–218.
9. Willis, J. and Robinson, J.E.1988. *Enterobacter sakazakii* meningitis in neonates. Infection Dis.J.,7:196-199.
10. Girlich, D.; Poirle, L.; Leelaporn, A.; Karim, A.; Tribuddharat, C.;

- Fennewald, M.; and Nordmann, P. 2001. Molecular epidemiology of the integrin-located VEB-1extended-spectrum  $\beta$ -lactamase isolates in Bangkok, Thailand. *J. Clin. Microbiol.*, 39, 175-182.
11. Kim, K.; Sik Jang, S.; Ki Kim, S.; Park, J.H.; Heu, S.; and Rye, S. 2008. Prevalence and genetic diversity of *Enterobacter sakazakii* in ingredients of infant foods. *Int. J. Food Microbiol.*, 122, 196-203.
  12. Stock, I.; and Wiedemann, B. 2002. Natural antibiotic susceptibility of *Enterobacter amnigenus*, *Enterobacter cancerogenus*, *Enterobacter gergoviae* and *Enterobacter sakazakii* strains. *Clin. Microbiol. and Infec.*, 8, 564-578.
  13. Hunter, C.J.; Petrosyan, M.; Ford, H.R.; and Prasadarao, N.V.2008. *Enterobacter sakazakii*: an emerging pathogen in infants and neonates. *Surg Infect (Larchmt.)*, 9(5):533–539.
  14. Shareef, A. Y.; Fathi, N. A.; and Noori, H. S. 2012. Detection of the ability of isolated *Enterobacter sakazakii* from powdered milk for enterotoxins production. *J.*12: 1-3.
  15. 1-3.
  16. Acker van, J.; de Smet, F.; Muyldermans, G.; Bougatef, A.; Naessens, A.; and Lauwers, S.2001.Outbreak of necrotizing enterocolitis associated with *Enterobacter sakazakii* in powdered milk formula. *J. Clin. Microbiol.*,39: 293–297.
  17. Restaino, L. E.W.; Frampton, W.C.; Lionberg, M.; and Becker, R.J. 2006. A chromogenic plating medium for the isolation and identification of *Enterobacter sakazakii* from foods, food ingredients, and environmental sources. *J. Food Prot.*, 69:315-322.
  18. Block, C.; Peleg, O.; Minster, N.; Bar-Oz, B.; Simhon, A.; Arad, I.; and Shapiro, M. 2002. Cluster of neonatal infections in Jerusalem due to unusual biochemical variant of *Enterobacter sakazakii*. *European J. Clin. Microbiol. Infect. Dis.*, 21:613–16.
  19. Quinn,P.J.; Carter, M.E.; Markey, B. and Carter, G.R.2004.Clinical veterinary microbiology.6<sup>th</sup> ed. Mosby an imp. Wolf, London.
  20. Iversen, C.; Druggan, P.; and Forsythe, S. 2004. A selective differential medium for *Enterobacter sakazakii*, a preliminary study. *Intl. J. Food Microbiol.*,96 (2): 133-139.
  21. Weiss, C.; Becker, B.; and Holzapfel, W. 2005. Application and acceptability of three commercial systems for detection of *Enterobacter sakazakii* in ready-to-eat vegetable salads [Einsatz und Eignung dreier kommerzieller Systeme zum Nachweis von *Enterobacter sakazakii* in verzehrsfertigem Mischsalat]. *Arch. Lebensmittelhyg.*, 56, 34–38.
  22. Oonaka, K.; Furuhashi, K.; Hara, M.; and Fukuyama, M.2010. Powder infant formula milk contaminated with *Enterobacter sakazakii*. *Jpn. J. Infect. Dis.*, 63: 103-107.
  23. Barron, C.J.; Hurrell, E.; Townsend, S.; Cheetham, P.;Locarrillo, C.; Fayet, O.; Prere, M.F.; and Forsythe, S.J. 2007. Genotypic and phenotypic analysis of *Enterobacter sakazakii* strains from an outbreak resulting in fatalities in a neonatal intensive care unit in France. *J. Clin. Microbiol.*, 45:3979-3985.
  24. Lampel, K.A.; and Chen, Y.2009. Method for the isolation and detection of *Enterobacter sakazakii* (*Cronobacter*) from powdered infant formula. *Int. J. Food Microbiol.*, 31;136(2):179-84.

25. Muytjens, H.L.; Roelofs-Willemse, H.; and Jaspar, G.H. 1988. Quality of powdered substitutes for breast milk with regard to members of the family Enterobacteriaceae. J. Clin. Microbiol. 26:743-6
26. Fu, S.; Gao, J.; Liu, Y.; and Chen, H. 2011. Isolation of *Cronobacter* spp. isolates from infant formulas and their survival in the production process of infant formula. Czech J. Food Sci., 29: 391–399.
27. Ranjan, K.D. 2007. Textbook of diagnostic microbiology. medical collage and hospital, medical publishers (p) Ltd Newdelhi., PP:124.
28. Farmer, J.J.; Asbury, M.A.; Hickman, F.W.; and Brenner, D.J. 1980. The *Enterobacteriaceae* Study Group. *Enterobacter sakazakii*, new species of *Enterobacteriaceae* isolated from clinical specimens. Intl. J. Sys. Bacteriol., 30: 569-84.
29. Block, C.; Peleg, O.; Minster, N.; Bar-Oz, B.; Simhon, A.; Arad, I.; and Shapiro, M. 2002. Cluster of neonatal infections in Jerusalem due to unusual biochemical variant of *Enterobacter sakazakii*. European J. Clin. Microbiol. Infect. Dis., 21:613–16.
30. Thompson, T. 2010. Food safety survey report Microbiological quality of infant formula. Government of South Australia Food Act Report Year ending 30 June.
31. Sani, N.A. and Yen Yi, L. 2011. Enterobacteriaceae, *Cronobacter (Enterobacter) sakazakii* and Microbial Population in Infant Formula Products in the Malaysian Market. Sains Malaysiana., 40(4):345–351.
32. Güner, A.; Doruer, Y.; Ali, C. M.; Yalçın, S.; Gülsen, S.; and Telli, N. 2011. An investigation on the prevalence of *Cronobacter sakazakii* in powdered infant formula consumed in Turkey. J. of Food, Agriculture and Environment., V.9 (2): 82 - 84.
33. Joosten, H. and Iversen, C. 2009. Development of a CEN-ISO horizontal standard method for detection of *Cronobacter*.
34. CAC. 2008. Codex Alimentarius Commission. Code of Hygienic Practices for Powdered Formulae for Infants and Young Children., CAC/RCP 66.
35. Steigerwalt, A.G.; Fanning, G.R.; Fife-Asbury, M.A.; and Brenner, D.J. (1979). DNA relatedness among species of *Enterobacter* and *Serratia*. Canad J. Microbiol. 22:121-37.
36. Campos, L.C.; Lobianco, L.F.; Seki, L.M.; Santos, R.M.; and Asensi, M. D. 2007. Outbreak of *Enterobacter hormaechei* septicaemia in newborns caused by contaminated parenteral nutrition in Brazil. J. Hosp. Infect., 66, 95–97.
37. Mezzatesta, M.L.; Gona, F.; and Stefani, S. 2012. *Enterobacter cloacae* Complex: Clinical Impact and Emerging Antibiotic Resistance Future Microbiol., 7(7):887-902.
38. Drudy, D.; O'Rourke, M.; Murphy, M.; Mullane, N.R.; O'Mahony, R.; and Kelly, L. 2006. Characterization of a collection of *Enterobacter sakazakii* isolates from environmental and food sources. Intl. J. Food Microbiol. 110 (2): 127-134.
39. Shadliya-Matug, M.; Aidoo, K.E.; Candlish, A.A.; and Elgerbi, A.M. 2008. Evaluation of some antibiotics against pathogenic bacteria isolated from infant foods in North Africa. Open Food Sci. J., 2:95–101.
40. Zhou, X.; Gao, J.; Huang, Y.; FU, S.; and Chen, H. 2011. Antibiotic resistance pattern of *Klebsiella pneumonia* and *Enterobacter sakazakii* isolates from powdered

- infant formula African J. of Microbiol., Research V. 5(19), pp. 3073-3077.
41. El-Sharoud, W.; O'Brien, S.; Negredo, C.; Iversen, C.; Fanning, S.; and Healy, B. 2009. Characterization of Cronobacter recovered from dried milk and related products. BMC Microbiol., 9: 9.
42. Pitout, J.D.; Moland, E.S.; Sanders, C.C.; Thomson, K.S.; and Fitzsimmons, S.R. 1997. Beta-lactamases and detection of beta-lactam resistance in *Enterobacter spp.* Antimicrob Agents Chemother., 41:35-39.
43. Al-Nabulsi, A.A.; Osaili, T.M.; Elabedeen, N.A.; Jaradat, Z.W.; Shaker, R.R.; Kheirallah, K.A.; Tarazi, Y.H.; and Holley, R.A. 2011. Impact of environmental stress desiccation, acidity, alkalinity, heat or cold on antibiotic susceptibility of *Cronobacter sakazakii*. Int.J.Food Microbiol., 30;146(2):137-43.

## التحري عن جرثومة *Cronobacter sakazakii* (*Entrobacter sakazakii*) في الحليب الباودر للرضع والحليب الخام في العراق

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### الخلاصة :

أجريت هذه الدراسة للكشف عن *C.sakazakii* من الحليب الباودر للرضع و الحليب الخام . تم جمع 200 عينة من الحليب الباودر من الرضع المصابين المرضي في المستشفى و الأسواق المحلية بالإضافة الى جمع 16 عينة من الحليب الخام . سجلت هذه الدراسة العزل الاول بكتريا *C.sakazakii* و *Enterobacter spp.* من الحليب الخام في العراق. توزعت *C.sakazakii* و *Enterobacter spp* بين العزلات باستخدام نظام فيتيك GN2 - أعطى 16/1 ( 6.25 % ) عزلة من *C.sakazakii* و 16/4 ( 25 % ) من عزلات *Enterobacter spp* *E.cloacae ssp. cloacae* and *E.cloacae ssp. dissolvens*, تشمل *Enterobacter spp* المعزولة تشمل *E.hormaechei*, and *E.ludwigii* الاختلافات بين النسب المئوية لكل عزل كانت غير معنوية (  $P < 0.05$ ). أظهرت نتائج المقاومة للمضادات الحيوية باستخدام نظام فيتيك - GN2 العزلات *C.sakazakii* مقاومة 100% cefazolin and cefoxitin ، وجد ان عزلات *C.sakazakii* عالية الحساسية ل كثير من المضادات الحيوية ويشمل ( Imipenem, Meropenem, Amikacin, Gentamicin, Tobramycin, Ciprofloxacin, Levofloxacin, Nitrofurantion, Trimethoprim \sulfamethoxazole, Ampicillin, Ampicillin\sulbactam, Piperacillin \Tazobactam, Ceftazidime, Ceftriaxone, Cefepime Azetreonom and augmentin ). لم يتم عزل *C.sakazakii* في جميع عينات الحليب الباودر الخاص بالرضع المتوفر في الأسواق المحلية.