

# **Antioxidant Capacity of Benalu Duku Leaves Alcoholic Extract on SOD Level and Pancreatic Cytology in Induced Diabetic Rats**

Anggun Syafitri<sup>1[,](mailto:dwirita@usu.ac.id)2</sup> DS, Yuandani<sup>1</sup> DS, Tri Widiyawati<sup>3</sup> DS, Dwi Rita Anggraini<sup>4</sup> DS, *Syukur Berkat Waruwu\* 5*

<sup>1</sup>Departement of Pharmacology, Faculty of Pharmacy, Universitas Sumatera Utara, Medan, Indonesia.

<sup>2</sup>Faculty of Pharmacy, Institut Kesehatan Deli Husada, Deli Serdang, Indonesia.

<sup>3</sup>Department of Pharmacology and Therapeutics, Faculty of Medicine, Universitas Sumatera Utara, Medan, Indonesia.

<sup>4</sup>Department of Anatomy, Faculty of Medicine, Universitas Sumatera Utara, Medan, Indonesia.

<sup>5</sup>Faculty of Pharmacy and Health Sciences, Universitas Sari Mutiara Indonesia, Medan, Indonesia.

\*Corresponding Author.

Received 21/05/2024, Revised 27/07/2024, Accepted 29/07/2024, Published Online First 20/12/2024

© 2022 The Author(s). Published by College of Science for Women, University of Baghdad. This is an Open Access article distributed under the terms of th[e Creative Commons Attribution 4.0 International License,](https://creativecommons.org/licenses/by/4.0/) which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

# **Abstract**

Diabetes mellitus causes damage to pancreatic β cells and oxidative stress due to an imbalance of oxidants and antioxidants in the body. Controlling hyperglycemia by administering conventional drugs and with long-term use carries the risk of side effects, so traditional treatment is recommended. Benalu Duku (*Dendophthoe pentandra* (L.) Miq) is a plant considered a parasite. However, it has the potential to be developed as a diabetes drug because it contains metabolites that can be used as drugs that come from nature. This study aims to test phytochemicals and examine the effect of ethanol extract of Benalu Duku leaves (EEBD) on superoxide dismutase (SOD) levels in streptozotocin-nicotinamide-induced diabetic white Wistar rats, blood glucose levels were also examined, as well as conducting histological analysis of pancreatic β cells. The results of the phytochemical examination showed that it contained alkaloids, flavonoids, glycosides, saponins, tannins, and triterpenoids. Research shows that giving EEBD for 28 days can significantly reduce blood glucose levels compared to the Na-CMC group. SOD levels also increased with respective values of  $30.97 \pm 0.84$ ,  $21.99 \pm 0.61$ ,  $30.52 \pm 1.30$ ,  $28.55 \pm 1.30$ ,  $28.99 \pm 0.95$ , and  $29.00 \pm 0.86$  pg/mL. Pancreatic histology also showed differences between qualitative and quantitative, indicating pancreatic repair and increased surface area of the islets of Langerhans. This plant has the potential to be developed into a new medicinal ingredient that comes from nature.

**Keywords:** *Dendrophthoe pentandra*, Diabetic, Hematoxylin-eosin, Langerhans, Pancreatic, Superoxide Dismutase.

# **Introduction**

Diabetes mellitus is a metabolic condition that can cause many long-term consequences, including nephropathy, neuropathy, retinopathy, and cardiomyopathy. Due to the accumulation of damage to several organs, this condition is sometimes known as a silent killer and is even a high-risk condition that can be  $fatal<sup>1,2</sup>$ . . Diabetes causes chronic hyperglycemia due to impaired insulin secretion due

to damage to pancreatic β-cells or impaired insulin receptor sensitivity<sup>3-5</sup>. Damage to pancreatic  $\beta$ -cells is often associated with oxidative stress due to an imbalance between oxidants and antioxidants in the body<sup>6-8</sup>. Persistent hyperglycemia causes increased production of free radicals, especially SOD. Increased glucose in diabetes results in the accumulation of ROS in pancreatic beta cells. This

accumulation of ROS will damage the cells where they are located. In type 2 diabetes, ROS reduces insulin synthesis and activates the beta cell apoptosis pathway<sup>9,10</sup>. Controlling hyperglycemia in diabetes mellitus patients has been controlled by administering oral antidiabetic agents. However, their use must be consumed for life, requiring high medical costs. Apart from that, long-term use poses a risk of severe side effects, so the use of traditional medicine is more recommended $11-13$ .

Benalu Duku (*Dendrophthoe pentandra* (L.) Miq) is a parasitic plant included in 3000 other plant species with medicinal potential. Because it is considered a parasite, this plant is often thrown away because it is thought to interfere with the growth of Duku plants.

### **Materials and Methods**

#### **Sample Collection**

Benalu Duku leaves were obtained from the Duku fields of Medan Johor Village, Medan, North Sumatra Province, Indonesia. Plant identification was carried out at the Medanense Herbarium, University of Sumatera Utara, Indonesia, with a number 136/MEDA/2022. Other materials used include nicotinamide (Brataco-Chem), streptozotocin (Brataco-Chem), natrium carboxymethyl cellulose 0.5% (Brataco-Chem), glibenclamide, ethanol 96%, distilled water, trichloroacetic acid 20%, thiobarbituric acid 0.67%, and enzyme-linked immunosorbent assay (ELISA) kit.

#### **Extract Preparation**

Five hundred grams of Benalu Duku leaf simplicia powder was put into a vessel then added with ethanol solvent until it was soaked with ten parts of ethanol solvent, covered and soaked for the first 6 hours, then left for 18 hours. The macerate is separated by filtering. The filtering process was repeated three times, using half the solvent volume in the first filtering. The extract was concentrated using a rotary evaporator at a temperature of  $\pm$  40<sup>o</sup>C until a thick extract was obtained. Phytochemical screening tests examine flavonoid, alkaloid, saponin, tannin, glycoside, and steroid/triterpenoid compounds<sup>22,23</sup>.

#### **Preparation of Test Animals**

The test animals used were male white Wistar rats with a body weight of 180-200 grams, divided into six groups, each consisting of 4 animals. This research procedure was carried out based on guidelines and approval from the Animal Research



However, leaves that have been considered parasitic benefit human health $14,15$ . The compounds that have been isolated from Benalu Duku leaves are Quercetin 3-methyl ether-7-O-arabinoside, Quercetin 3- oarabinoside-7-o-glucoside, and Quercetin 3-O-α-Rhamnoside, each of which has different activities. High as an antioxidant, antibacterial, and<br>antidiabetic<sup>16-18</sup> A flavonoid molecule called A flavonoid molecule called quercetin 3-methyl ether-7-O-arabinoside can be a free radical neutralizer, protecting pancreatic beta cells from harm and promoting insulin release<sup>19-21</sup>. This research was conducted to test the effect of ethanol extract of Benalu Duku leaves on SOD levels and the protective effect on pancreatic β cells in diabetic rats induced by streptozotocin-nicotinamide.

Ethics Committees (AREC), Department of Biology, Faculty of Mathematics and Natural Sciences, University of Sumatera Utara, Indonesia, with approval number 0795/KEPH-FMIPA/2022. The rats were separated into six groups, each consisting of 4 rats, as follows:

Group 1: Normal

Group 2: Na-CMC (natrium carboxymethyl cellulose) 0.5%

Group 3: Glibenclamide 0.45 mg/kg BW

Group 4: EEBD (ethanol extract of benalu duku leaves) 100 mg/kg BW

Group 5: EEBD 200 mg/kg BW

Group 6: EEBD 400 mg/kg BW

Before the test, the rats were first acclimatized to the laboratory environment for a week, ensuring their comfort and reducing stress. They were handled carefully without causing fear. The night before treatment, the animals were fasted first. After fasting for 18 hours, rats were induced by administering 230 mg/kg BW nicotinamide solution intraperitoneally. After 15 minutes, continue with administration of 65 mg/kg BW streptozotocin solution intraperitoneally. Blood glucose levels were measured using a glucometer (Easy Touch® GCU) after 72 hours of induction. Rats with fasting blood glucose levels >200 mg/dL are said to be diabetic and can be used in testing. The extract was given orally every day for 28 days starting when the rats had diabetes $24.25$ .

# **Measurement of SOD Levels**

On the last day, the rats were sacrificed using anesthesia. Blood was taken (3 - 3.5 mL) from the



heart to collect plasma serum, which was used to measure SOD levels using the ELISA method. The absorbance was read with a microplate reader at a wavelength of 450 nm, and the levels were calculated $26,27$ .

### **Histological Observations**

The rat pancreas was analyzed histologically using the hematoxylin-eosin staining method<sup>28</sup>. Then, observations can be made under a microscope with  $400x$  magnification<sup>29</sup>. The structure of the pancreatic

### **Results and Discussion**

#### **Phytochemical Screening**

The results of phytochemical screening of the EEBD were obtained to obtain information on the classes of secondary metabolite compounds. Phytochemical examination showed the presence of alkaloids, flavonoids, glycosides, saponins, tannins, and triterpenoid compounds. The results of phytochemical screening can be seen in Fig. 1. Phytochemical examination showed the presence of alkaloid compounds. When the Mayer reagent solution was added, it formed a white lumpy precipitate; with the Bouchardat reagent solution, a blackish brown precipitate was formed, and the Dragendorff reagent solution formed a red color. Examining the flavonoid compound group with

islands of Langerhans is examined by observing the islands of cells buried in the exocrine tissue of the pancreas. Then, the examination continues by calculating the area of the pancreatic islands of Langerhans $30$ .

#### **Data Analysis**

Statistical analysis used Anova and the Post Hoc Tukey HSD test to determine whether there were fundamental differences between treatments<sup>31</sup>.

magnesium powder and concentrated hydrochloric acid produces a red-colored solution. Examination of the group of glycoside compounds by adding Molisch's reagent and concentrated sulfuric acid forms purple rings. The sample was added with hot distilled water and shaken vigorously to produce stable foam, then 2 N HCl was added, indicating the presence of the saponin group of compounds. Adding FeCl<sub>3</sub> gives a blackish-green color, indicating the presence of tannin compounds. Examination of the triterpenoid/steroid compound group by adding a few drops of Liebermann-Burchard reagent produces a pink or purple color, which indicates the triterpenoid compound group<sup>32,33</sup>.



**Figure 1. Results of phytochemical screening of EEBD**

### **Blood Glucose Level Examination Results**

Glucose was checked 72 hours after induction, and glucose was rechecked on the last day of

treatment. The examination results are displayed in Table 1.





**Note:** Data is presented in the form of average and standard errors. <sup>a</sup>: significantly different from the Na-CMC control group, <sup>b</sup>: significantly different from the glibenclamide group,  $\degree$ : significantly different from the normal group.

Blood glucose levels in induced rats were high on average except in the normal group (not induced). This shows that the rats used for the experiment were in a state of hyperglycemia. Streptozocinnicotinamide induction can increase blood glucose levels<sup>34</sup>. On the last day, blood glucose levels decreased in all treatment groups. However, the Na-CMC group did not experience a significant decrease.

#### **SOD Level Examination Results**

The results of examining SOD levels, as shown in Table 1.





**Note:** Data is presented in the form of average and standard errors. <sup>a</sup>: significantly different from the Na-CMC control group,  $b$ : significantly different from the glibenclamide group, <sup>c</sup>: significantly different from the normal group.

The average SOD concentration value in the Na-CMC group had the lowest concentration. Administration of nicotinamide and streptozotocin as triggers of oxidative stress will reduce antioxidant resistance in the body by showing a decrease in SOD levels. Administration of nicotinamide may influence the SOD levels examined, where nicotinamide is a direct precursor of NAD<sup>+</sup> and an inhibitor of poly ADP ribose, which has the effect of increasing ATP levels in cells, thereby reducing cell damage<sup>35-37</sup>.

Administration of EEBD had a significant effect on increasing SOD concentrations. The three EEBD groups showed significant differences with the Na-CMC group. The chemical compounds such as flavonoids, tannins, and saponins from various types of plants are known to have antioxidant effects<sup>38,39</sup>. The low SOD levels in the Na-CMC group indicate the high oxidative stress that occurs in diabetic rats without therapy. Diabetes mellitus increases ROS, thereby exacerbating oxidative stress. Oxidative stress causes an imbalance in the system for the formation and capture of free radicals, thereby reducing antioxidant activity $40-42$ .

#### **Histological Examination on Pancreatic**

Histological observations on pancreatic preparations from normal groups and treated groups showed different structures, as in Fig. 2. Based on the results of the observations, the normal group showed pancreatic features, such as pancreatic acinar cells and Langerhans cells. Langerhans cells appear with regular cell membranes; alpha cells are located at the edge, with small, dense, round nuclei and little cytoplasm (red stars), while beta cells are located in the center with larger, brighter nuclei, eosinophilic cytoplasm (yellow stars). In the positive control, Langerhans cells appeared with a large surface area with many cells as in the normal group, and the cell membrane was still regular (black arrow). Meanwhile, in the Na-CMC group, Langerhans cells appeared with a smaller surface area and fewer cells. The cells in the central (middle) section appear to be experiencing edema (hydrophilic degeneration) marked with a yellow star. In the 100 mg/Kg BW group, Langerhans cells showed a smaller surface area with fewer cells and irregular cell membranes (black arrows). The 200 mg/KgBW group showed Langerhans cells with a larger surface area than the normal group and a more significant number of cells. It can be seen that the cells in the central

(middle) section are experiencing proliferation, marked by yellow stars and slightly irregular cell membranes (black arrows). Finally, in the 400 mg/Kg BW group, Langerhans cells appeared with a larger surface area and a more significant number of cells resembling the normal group. The cell membrane was slightly irregular (black arrow). Induction of diabetes with streptozotocin and nicotinamide causes pancreatic beta-cell necrosis<sup>43</sup>. Beta cells show a significant decrease in moderate diabetes, whereas, in severe diabetes, beta cells are not even found. Langerhans cells with a smaller surface area and a smaller number of cells in the Na-CMC group indicate damage to the pancreatic tissue; the cells in the central part also experience hydrophic degeneration. Hydrophilic degeneration indicates that reversible changes have occurred in the cells, so if the toxic exposure is stopped, the damaged cells will return to normal. Continued degeneration will cause cell death. Liver cell death causes hepatocytes to be unable to return to their normal form (irreversible). Hydrophilic degeneration increases intracellular water content, which causes the cytoplasm and organelles to swell and form vacuoles. Damage to the cell membrane's permeability causes obstruction to sodium flow out of the cell, causing excessive ions and water to enter the cell. A lack of oxygen, calcium deficiency, severe shock, and diabetes mellitus can cause this hydrophilic degeneration<sup>44,45</sup>.



**Figure 2. Pancreatic histopathology (400x), A: Normal group; B: Glibenclamide; C: Na-CMC; D: EEBD 100 mg; E: EEBD 200 mg; F: EEBD 400 mg; IL: Islet of pancreas; PA: Panceatic acinar; \*: alpha cells, \*: beta cells;: cell membrane**

Quantitative calculations of Langerhans islands were conducted to assess improvements. The results are shown in Fig. 3. The area of the islets of Langerhans was highest in the normal group, significantly different from the Na-CMC group. The EEBD 200 mg/kg BW group had a lower mean area, while the highest was observed at the EEBD dose of 400 mg/kg BW. These findings suggest the potential for beneficial pharmacological effects of EEBD. In diabetes mellitus sufferers, Langerhans Islands undergo morphological changes in both number and size. The islets of Langerhans, a collection of endocrine glands spread across the pancreas, resemble islands with numerous blood capillaries passing through them<sup>46-48</sup>.

### **Conclusion**

EEBD has potential sound pharmacological effects that can significantly increase the SOD levels of



**Figure 3. Results of calculating the area of the pancreatic islets of Langerhans. Note: Data is presented in the form of average and standard errors**

streptozotocin-nicotinamide-induced diabetic rats. On Langerhans Island, EEBD increased the

Langerhans surface area of diabetic rats compared to Na-CMC group. The leaves of this plant have the potential to be developed in the future and analyzed

# **Acknowledgment**

The authors would like to sincerely thank the Master of Pharmacy Study Program at the Faculty of

# **Authors' Declaration**

- Conflicts of Interest: None.
- We hereby confirm that all the figures and tables in the manuscript are ours. Furthermore, figures and images, that are not ours, have been included with the necessary permission for re-publication, which is attached to the manuscript.
- The author has signed an animal welfare statement.

#### **Authors' Contribution Statement**

This manuscript was created in collaboration with all authors: Y. conceptualized and designed the research, A.S. collected samples, performed analysis, and wrote the manuscript, D.R.A. performed histological analysis, T.W. the result

#### **References**

- 1. Mauricio D, Alonso N, Gratacòs M. Chronic diabetes complications: the need to move beyond classical concepts. Trends Endocrinol Metab. 2020; 31(4): 287- 295. <https://doi.org/10.1016/j.tem.2020.01.007>
- 2. Alam S, Hasan M K, Neaz S, Hussain N, Hossain M F, Rahman T. Diabetes mellitus: insights from epidemiology, biochemistry, risk factors, diagnosis, complications and comprehensive management. Diabetology. 2021; 2(2): 36-50. <https://doi.org/10.3390/diabetology2020004>
- 3. Mukai E, Fujimoto S, Inagaki N. Role of reactive oxygen species in glucose metabolism disorder in diabetic pancreatic β-cells. Biomolecules. 2022; 12(9): 1-15.<https://doi.org/10.3390/biom12091228>
- 4. Abed B A, Farhan L O, Dawood A S. Relationship between serum nesfatin-1, adiponectin, resistin concentration, and obesity with type 2 diabetes mellitus. Baghdad Sci J. 2023; 21(1): 117-123. <https://doi.org/10.21123/bsj.2023.8119>
- 5. Jabbar A A, Abdulrahman K K, Abdulsamad P, Mojarrad S, Mehmetçik G, Sardar A S. Phytochemical profile, antioxidant, enzyme inhibitory and acute toxicity activity of Astragalus bruguieri. Baghdad Sci J. 2023; 20(1): 157-165. <https://doi.org/10.21123/bsj.2022.6769>

regarding the active ingredients contained in this plant.

Pharmacy, University of Sumatera Utara, Medan, Indonesia.

- No human studies are present in the manuscript.
- Ethical Clearance: The project was approved by the local ethical committee at Animal Research Ethics Committees (AREC), Department of Biology, Faculty of Mathematics and Natural Sciences, University of Sumatera Utara, with approval number 0795/KEPH-FMIPA/2022.

interpretation, S.B.W. revisions, proofreading, edit the manuscript with revisions and now take care of publishing. All authors read and approved the final manuscript.

- 6. Anastasiou I A, Eleftheriadou I, Tentolouris A, Koliaki C, Kosta O A, Tentolouris N. The effect of oxidative stress and antioxidant therapies on pancreatic β-cell dysfunction: results from in vitro and in vivo studies. Curr Med Chem. 2021; 28(7): 1328-1346. [https://doi.org/10.2174/092986732766620052613564](https://doi.org/10.2174/0929867327666200526135642) [2](https://doi.org/10.2174/0929867327666200526135642)
- 7. Dinić S, Arambašić Jovanović J, Uskoković A, Mihailović M, Grdović N, Tolić A, et al. Oxidative stress-mediated beta cell death and dysfunction as a target for diabetes management. Front Endocrinol (Lausanne). 2022; 13: 1-20. <https://doi.org/10.3389/fendo.2022.1006376>
- 8. Al-Chalabi N S, Al-Sawaf R N. Effect of polyherbsmixture composed of nigella sativa, trigonella foenum - graceum, cyperus rotundus and teucrium polium on the levels of malondialdehyde and glutathione for diabetic patients type II. Baghdad Sci J. 2013; 10(3): 854–865. [https://doi.org/10.21123/bsj.2013.10.3.854-](https://doi.org/10.21123/bsj.2013.10.3.854-865) [865](https://doi.org/10.21123/bsj.2013.10.3.854-865)
- 9. Promyos N, Phienluphon P P, Wechjakwen N, Lainampetch J, Prangthip P, Kwanbunjan K. Inverse correlation of superoxide dismutase and catalase with type 2 diabetes among rural thais. Nutrients. 2023; 15(9): 1-14. <https://doi.org/10.3390/nu15092071>





10. Khin P P, Lee J H, Jun H S. Pancreatic beta-cell dysfunction in type 2 diabetes. Eur J Inflamm. 2023; 21: 1-13.

<https://doi.org/10.1177/1721727X231154152>

- 11. Heshmat R, Darvishi A, Abdi Dezfouli R, Nikkhah A, Radmanesh R, Moslemi E. A short-term economic evaluation of early insulin therapy compared to oral anti-diabetic drugs in order to reduce the major adverse events in type 2 diabetes patients in Iran. Curr Med Res Opin. 2024: 1-8. <https://doi.org/10.1080/03007995.2024.2333425>
- 12.Chaudhury A, Duvoor C, Reddy Dendi VS, Kraleti S, Chada A, Ravilla R, et al. Clinical review of antidiabetic drugs: implications for type 2 diabetes mellitus management. Front Endocrinol*.* 2017; 8: 1- 12.<https://doi.org/10.3389/fendo.2017.00006>
- 13.Blahova J, Martiniakova M, Babikova M, Kovacova V, Mondockova V, Omelka R. Pharmaceutical drugs and natural therapeutic products for the treatment of type 2 diabetes mellitus. Pharmaceuticals. 2021; 14(8): 1-32.<https://doi.org/10.3390/ph14080806>
- 14. Mochamad L, Malarvili S, Jasmine K, Lim V. In vitro analysis of quercetin-like compounds from mistletoe Dendrophthoe pentandra (L.) Miq as a potential antiviral agent for Newcastle disease. F1000Res. 2024; 12: 1-38. <https://doi.org/10.12688/f1000research.133489.5>

15. Awang M A, Nik Mat Daud N N N, Mohd Ismail N I, Abdullah F I, Benjamin M A Z. A review of Dendrophthoe pentandra (Mistletoe): phytomorphology, extraction techniques, phytochemicals, and biological activities. Processes. 2023; 11(8): 1-19[. https://doi.org/10.3390/pr11082348](https://doi.org/10.3390/pr11082348)

- 16. Kong D, Wang L, Niu Y, Cheng L, Sang B, Wang D, et al. Dendrophthoe falcata (L.f.) Ettingsh. and Dendrophthoe pentandra (L.) Miq.: A review of traditional medical uses, phytochemistry, pharmacology, toxicity, and applications. Front Pharmacol. 2023; 14: 1-18. <https://doi.org/10.3389/fphar.2023.1096379>
- 17. Hardiyanti R, Marpaung L, Adnyana I K, Simanjuntak P. Biochemical evaluation of Duku's mistletoe leave (Dendrophthoepentandra (L.) Miq) extract with antidiabetic potential. Rasayan J Chem*.* 2019; 12(03): 1569-1574.

<https://doi.org/10.31788/RJC.2019.1235272>

18. Hardiyanti R, Marpaung L, Adnyana I K, Simanjuntak P. Isolation of quercitrin from Dendrophthoe pentandra (L.) Miq leaves and it's antioxidant and antibacterial activities. Rasayan J Chem*.* 2019; 12(04): 1822-1827.

<https://doi.org/10.31788/RJC.2019.1235353>

- 19. Fahim M D, Rahman I, Naseem N, Imam N, Younus H, Ahsan H, et al. Antidiabetic potential of natural phytochemical antioxidants. JCNB. 2022; 3(2): 26-43. <https://doi.org/10.48185/jcnb.v3i2.610>
- 20. Mohamed G A, Omar A M, El-Araby M E, Mass S, Ibrahim S R M. Assessments of alpha-amylase

inhibitory potential of tagetes flavonoids through in vitro, molecular docking, and molecular dynamics simulation studies. Int J Mol Sci. 2023; 24(12): 1-22. <https://doi.org/10.3390/ijms241210195>

- 21. AL-Ishaq, Abotaleb, Kubatka, Kajo, Büsselberg. Flavonoids and their anti-diabetic effects: cellular mechanisms and effects to improve blood sugar levels. Biomolecules. 2019; 9(9): 1-35. <https://doi.org/10.3390/biom9090430>
- 22. Dalimunthe A, Muhammad M, Waruwu S B, Rafi M, Kaban V E, Satria D. Phytochemicals and proximate analysis of Litsea Cubeba Lour. barks. IOP Conf Ser Earth Environ Sci. 2023; 1188(1): 1-7. <https://doi.org/10.1088/1755-1315/1188/1/012012>
- 23. Dalimunthe A, Pertiwi D, Muhmmad M, Kaban V E, Nasri N, Satria D. The effect of extraction methods towards antioxidant activity of ethanol extract of Picria fel-terrae Lour. Herbs. IOP Conf Ser Earth Environ Sci. 2022; 1115(1): 1-6. [https://doi.org/10.1088/1755-](https://doi.org/10.1088/1755-1315/1115/1/012040) [1315/1115/1/012040](https://doi.org/10.1088/1755-1315/1115/1/012040)
- 24. Pottathil S, Nain P, Morsy M A, Kaur J, Al-Dhubiab B E, Jaiswal S, et al. Mechanisms of antidiabetic activity of methanolic extract of Punica granatum leaves in nicotinamide/streptozotocin-induced type 2 diabetes in rats. Plants. 2020; 9(11): 1-15. <https://doi.org/10.3390/plants9111609>
- 25. Widyawati T, Yusoff N A, Bello I, Asmawi M Z, Ahmad M. Bioactivity-guided fractionation and identification of antidiabetic compound of Syzygium polyanthum (Wight.)'s leaf extract in streptozotocininduced diabetic rat model. Molecules. 2022; 27(20): 6814-6829.

<https://doi.org/10.3390/molecules27206814> 26. Dalimunthe A, Satria D, Sitorus P, Harahap U, Angela I F D, Waruwu S B. Cardioprotective effect of hydroalcohol extract of andaliman (Zanthoxylum acanthopodium DC.) fruits on doxorubicin-induced rats. Pharmaceuticals. 2024; 17(3): 359-463.

- <https://doi.org/10.3390/ph17030359> 27. Satria D, Octora D D, Muhammad M, Rosidah, Silalahi J, Waruwu S B. In vivo analysis of Saurauia vulcani Korth. leaves extract as antihypercholesterolemic. Res J Pharm Technol*.* 2024; 17(5): 2051-2055. [https://doi.org/10.52711/0974-](https://doi.org/10.52711/0974-360X.2024.00325) [360X.2024.00325](https://doi.org/10.52711/0974-360X.2024.00325)
- 28. Abbas A K, Abbas N K, Ali R M, Abbas L K. Histological and biochemical parameters follow-up in experimental rats administrated dexamethasone and treated with green synthesis titanium dioxide nanoparticles using (Camillia sciences) extracts. Baghdad Sci J. 2020; 17(2): 663-669. [https://doi.org/10.21123/bsj.2020.17.2\(SI\).0663](https://doi.org/10.21123/bsj.2020.17.2(SI).0663)
- 29. Surbakti C, Sitorus P, Rosidah R, Satria D. Effect of Saurauia vulcani Korth. leaves on superoxide dismutase, HbA1c levels and insulin expression in hyperglycemic rats. Open Access Maced J Med Sci. 2019; 7(22): 3741-3744. <https://doi.org/10.3889/oamjms.2019.494>



- 30. Anggraini D R, Widyawati T, Syarifah S, Wahyuni A S. Evaluation of blood glucose level and microscopic pancreatic islets of langerhans treated with Lawsonia Inermis Linnaeus leaves ethyl acetate extract in streptozotocin-induced diabetic rat. Scitepress; 2018: 108-11[2.https://doi.org/10.5220/0010039101080112](https://doi.org/10.5220/0010039101080112)
- 31. Satria D, Sitorus P, Dalimunthe A, Waruwu S B, Asfianti V. Oral acute toxicity study of ethanol extract of Mobe leaves (Artocarpus lacucha Buch-Ham) in Wistar rats. Pharmacia. 2024; 71: 1-8. <https://doi.org/10.3897/pharmacia.71.e117500>
- 32.Rusdiana R, Widyawati T, Sari D K, Widjaja S S. Phytochemical analysis of the ethanol extract of Binahong (Anredera cordifolia (Ten.) Steenis) leaves by UV-Vis spectroscopy. Baghdad SciJ. 2024; 21(11): 3446-345[1.https://doi.org/10.21123/bsj.2024.9354](https://doi.org/10.21123/bsj.2024.9354)
- 33.Robiatun R R, Pangondian A, Paramitha R, Zulmai Rani, Gultom E D. Formulation and evaluation of hand sanitizer gel from clove flower extract (Eugenia aromatica L.). IJSTM. 2022; 3(2): 484-491. <https://doi.org/10.46729/ijstm.v3i2.472>
- 34. Lesa K N, Ahmad N, Mayangsari Y, Cahyanto M N, Saputra W D. Anti-diabetic effect of Okara Noodles on streptozotocin-nicotinamide induced diabetic rats. Trends Sci. 2024; 21(5): 7428-7430. <https://doi.org/10.48048/tis.2024.7428>
- 35. Ghasemi A, Khalifi S, Jedi S. Streptozotocinnicotinamide-induced rat model of type 2 diabetes (review). Acta Physiol Hung. 2014; 101(4): 408-420. <https://doi.org/10.1556/APhysiol.101.2014.4.2>
- 36. Margaritis I, Angelopoulou K, Lavrentiadou S, Mavrovouniotis I C, Tsantarliotou M, Taitzoglou I, et al. Effect of crocin on antioxidant gene expression, fibrinolytic parameters, redox status and blood biochemistry in nicotinamide-streptozotocin-induced diabetic rats. J Biol Res (Thessalon). 2020; 27(1):4-19. <https://doi.org/10.1186/s40709-020-00114-5>
- 37. Song B R, Alam M B, Lee S H. Terpenoid-rich extract of Dillenia indica L. bark displays antidiabetic action in insulin-resistant C2C12 cells and STZ-induced diabetic mice by attenuation of oxidative stress. Antioxidants. 2022; 11(7): 1227-1236[.https://doi.org/10.3390/antiox11071227](https://doi.org/10.3390/antiox11071227)
- 38. Putra E D L, Cintya H, Satria D. Antibacterial and antioxidant activities of ethanol extract of sukun *(Artocarpus altilis.)* leaves against *Pseudomonas*

*aeruginosa*. E3S Web Conf. 2021; 332: 1-6. <https://doi.org/10.1051/e3sconf/202133208006>

- 39. Satria D, Dalimunthe A, Pertiwi D, Muhammad M, Kaban VE, Nasri N, et al. Phytochemicals, proximate composition, minerals and volatile oil analysis of Zanthoxylum acanthopodium DC. fruits. F1000Res. 2023; 12: 227-230. <https://doi.org/10.12688/f1000research.128941.1>
- 40.Black H S. A synopsis of the associations of oxidative stress, ROS, and antioxidants with diabetes mellitus. Antioxidants. 2022; 11(10): 2003-2019. <https://doi.org/10.3390/antiox11102003>
- 41. Li Y, Ren K. The mechanism of contrast-induced acute kidney injury and its association with diabetes mellitus. Contrast Media Mol Imaging. 2020; 2020: 1- 10.<https://doi.org/10.1155/2020/3295176>
- 42. Yaribeygi H, Sathyapalan T, Atkin S L, Sahebkar A. Molecular mechanisms linking oxidative stress and diabetes mellitus. Oxid Med Cell Longev. 2020; 2020: 1-13.<https://doi.org/10.1155/2020/8609213>
- 43.Jaishree V, Narsimha S. Swertiamarin and quercetin combination ameliorates hyperglycemia, hyperlipidemia and oxidative stress in streptozotocininduced type 2 diabetes mellitus in wistar rats. Biomed Pharmacother. 2020; 130: 1-8. <https://doi.org/10.1016/j.biopha.2020.110561>
- 44. Nandi A, Yan L J, Jana C K, Das N. Role of catalase in oxidative stress- and age-associated degenerative diseases. Oxid Med Cell Longev. 2019; 2019: 1-19. <https://doi.org/10.1155/2019/9613090>
- 45. Ukratalo A M, Kaihena M, Sirait D P O, Pattimura N, Manery D E. Potential bark Cinnamomum burmanii in regenerating damaged liver cells of mice (Mus musculus) diabetes mellitus model. PCJN. 2023; 2(01): 30-40.<https://doi.org/10.58549/pcjn.v2i01.58>
- 46. Vargas-Soria M, García-Alloza M, Corraliza-Gómez M. Effects of diabetes on microglial physiology: a systematic review of in vitro, preclinical and clinical studies. J Neuroinflammation. 2023; 20(1): 57- 87[.https://doi.org/10.1186/s12974-023-02740-x](https://doi.org/10.1186/s12974-023-02740-x)
- 47. Adams M T, Blum B. Determinants and dynamics of pancreatic islet architecture. Islets. 2022; 14(1): 82- 100.<https://doi.org/10.1080/19382014.2022.2030649>
- 48. Slak Rupnik M, Hara M. Local dialogues between the endocrine and exocrine cells in the pancreas. Diabetes. 2024; 73(4): 533-541. [https://doi.org/10.2337/db23-](https://doi.org/10.2337/db23-0760) [0760](https://doi.org/10.2337/db23-0760)





# **القدرة المضادة لألكسدة للمستخلص الكحولي ألوراق بينالو دوكو على مستوى SOD وعلم خاليا البنكرياس في الجرذان المصابة بداء السكري**

**5 ، سيوكور بيركات واروو <sup>4</sup> ، دوي ريتا أنغرايني <sup>3</sup> ، تري ويدياواتي <sup>2</sup> ، يوانداني 2،1 أنغون سيافيتري** 

 قسم الصيدلة، كلية الصيدلة، جامعة سومطرة أوتارا، ميدان، إندونيسيا. كلية الصيدلة، معهد كيسهاتان ديلي هوسادا، ديلي سيردانغ، إندونيسيا. قسم الصيدلة والعالج، كلية الطب، جامعة سومطرة أوتارا، ميدان، إندونيسيا. قسم التشريح، كلية الطب، جامعة سومطرة أوتارا، ميدان، إندونيسيا. كلية الصيدلة والعلوم الصحية، جامعة ساري موتيارا إندونيسيا، ميدان، إندونيسيا.

# **الخالصة**

يتسبب داء السكري في تلف خاليا البنكرياس واإلجهاد التأكسدي بسبب عدم توازن المواد المؤكسدة ومضادات األكسدة في الجسم. إن السيطرة على ارتفاع السكر في الدم عن طريق إعطاء الأدوية التقليدية والاستخدام طويل الأمد ينطوي على مخاطر الآثار الجانبية، لذلك يوصى بالعالج التقليدي. بينالو دوكو ) (Miq) .L (*pentandra Dendophthoe*هو نبات يعتبر من النباتات الطبيعية. ومع ذلك، فمن الممكن تطويره كدواء لمرض السكري ألنه يحتوي على مستقلبات يمكن استخدامها كأدوية تأتي من الطبيعة. تهدف هذه الدراسة إلى اختبار المواد الكيميائية النباتية ودراسة تأثير المستخلص الإيثانولي لأوراق بينالو دوكو ( (EEBDعلى مستويات فوق أكسيد ديسموتاز ) (SODفي فئران ويستار البيضاء المصابة بداء السكري المستحث بالستربتوزوتوسين والنيكوتيناميد، كما تم فحص مستويات الجلوكوز في الدم، باإلضافة إلى إجراء التحليل النسيجي. تحليل خاليا البنكرياس. وأظهرت نتائج الفحص الكيميائي النباتي أنها تحتوي على قلويدات وفالفونيدات وجليكوسيدات وصابونين وعفص وترايتيربينويدات. تظهر األبحاث أن إعطاء EEBD لمدة 82 يو ًما يمكن أن يقلل بشكل كبير من مستويات الجلوكوز في الدم مقارنة بمجموعة .CMC-Naزادت مستويات SOD أي ًضا بقيم 79.03 ± 9.20 و 89.00 ± 9..9 و 79.08 ± 9.79 و 82.00 ± 9.79 و 82.00 ± 9.00 و 80.99 ± 9.2. بيكوغرام / مل. وأظهرت أنسجة البنكرياس أيضًا اختلافات بين النوعية والكمية، مما يشير إلى إصلاح البنكرياس وزيادة مساحة سطح جزر لانجرهانس يتمتع هذا النبات بإمكانية تطويره ليصبح مكونًا طبيًا جديدًا يأتي من الطبيعة.

**الكلمات المفتاحية:** *pentandra Dendrophthoe*، السكري، الهيماتوكسيلين أيوزين، النجرهانز، البنكرياس، سوبر أكسيد ديسموتاز.