DOI: http://dx.doi.org/10.21123/bsj.2017.14.2.0356

Production and Extraction of Siderophores-Catecholatefrom -MDR-Acinetobacter baumannii

Assist. Prof. Dr. Intesar N. Khelkal

Department of Biology, College of Science, Al-Mustansiriya University, Baghdad, Iraq.

Received 14/8/2016 Accepted 9/11/2016

This work is licensed under a Creative Commons Attribution 4.0 International License

Abstract:

Siderophores are low molecular weight organic compounds produced by microorganisms growing under low iron concentration. In this study we describe the detection, production and extraction of siderophores secreted by *Acinetobacter baumannii* (Multiple-drug resistant) pathogens.

One hundered twenty Gram –negative non lactose fermenter bacilli isolates have been collected from three hospitals at Baghdad city over three months. Primary identification of these isolates is performed by standard diagnostic methods (biochemical tests and API 20 NE); 19 clinical isolates of *A. baumannii* are cultured on CHROMagar (highly selective medium for detection of MDR Acinetobacter) as well as diagnoses is documented by using Vitek 2 system. Isolates are examined towards 11 different antibiotics. High resistance is recognized for most isolates. Detection of siderophore has been done by examining the isolates on M9 minimum medium; 5 isolates (26%) are producers for siderophore, the highest producing one is isolated from sputum and chosen to extract siderophore catecholate . (Ab5S) isolate is examined on specific synthetic medium for production then siderophore molecules are extracted by ethyl acetate .Weight of dried extract is determined (115 mg/ml) and siderophore chemical nature has been assessed which appeared as catecholate.

Key words: A. baumannii , Siderophores, Catecholate

Introduction:

Siderophores are low molecular weight (<14 kDa) iron chelating compounds synthesized in large quantity under iron limitation conditions. There are three major types of siderophores; hydroxamate, catecholate and carboxylate [1]. Iron is a necessary element for the growth of bacterial cells because it acts as a catalyst in enzymatic processes, electron transfer, DNA, RNA syntheses and oxygen metabolism [2]. Iron is also essential for biofilm production because it stabilizes the polysaccharide layer and arranges surface motility [3].

One of the most important steps in initiating an infection is the availability of iron [4]. There are different methods to acquire iron by microorganisms; production of siderophores represents the first and more important method, iron adheres to the bacterial cell by specific receptors and moves inside by common transport techniques [5].

Genes of siderophore biosynthesis are responsible for bacterial infection in mouse, they activate exotoxins formation, affects cell movement and biofilms maturity [6].

Few studies show the ability of clinical isolates of *A.baumannii* to grow and produce siderophore compounds under iron-deficient condition [7].

A novel siderophore, called acinetobactin, with both catecholate and hydroxamate functional groups are isolated from low-iron cultures of *A*. *baumannii* ATCC 19606 [8].

Aim of the study: Siderophores have many medical applications, the most important one when act as avechile for transport antibiotics inside bacterial cell so extraction of these molecules should be investigated.

Materials and Methods:

1-Bacterial Isolates : One hundered twenty specimens belonging to non fermenter Gram-negative bacilli have been collected from three hospitals at Baghdad city over three months, 19 clinical isolates have been identified as *A. baumannii*, the diagnosis is confirmed by using highly selective medium CHROMagar Acinetobacter and Vitek 2 system.

2-Antibiotic Susceptibility: *A. baumannii* isolates are tested against 11 different antibiotic discs which have been provided by Bioanalyse (Turkey). **3-Detection of Siderophore**: All the isolates are cultured on M9 minimum solid medium which is prepared according to [9] Sambrook *et al.*,(1989) and modified by [10] Shenker *et al.*,(1992) as follows:

A-Dissolving : Na_2HPO_4 (6g) ; KH_2PO_4 (3g) ;NaCl (0.5g) ; NH_4Cl (1g) ; agaragar(15g) in one liter D.W,adjusted pH to 7.2, autoclaved,cooled to 45°C.

B- The following components are added to the medium prepared in (A):

20ml of MgSo4 (0.5g/20ml); 1ml Dipyridine (0.005g/10ml); 1ml CaCl₂ (0.03g/10 ml); 10 ml Glucose (2g/10ml). The components are sterilized by filteration using 0.22μ M millipore filters. The medium is then supplemented with 0.1g thiamine.

C- The components are well mixed and poured in disposable sterile plates,.

D-After being solidified, the plates are inoculated with tested isolates (touch by sterile woody stick) and incubated at 37°C for 24 hrs.

E- If the isolate is siderophore producing, the growth will appear as small, single and seperated colonies on M9 medium [8].

4-Siderophore production: A synthetic medium with the following components per liter is used:

mannitol	10 g
sodium gluconate	2 g
K ₂ HPO ₄	0.5g
$MgSO_4$	0.2g
NaCl	0.1g

pH was adjusted to 7 and autoclaved [11].

In order to avoid iron contamination , inoculation of the of producer isolate is performed by sterile woody stick and incubating the culture for 20hrs. at 35° C.

Note: All the flasks and glassware materials are soaked with acid, rinsed several times with water before using to minimize iron concentration (8).

5-Extraction of siderophore: According to the method of Jadhav and Desai (1992); bacterial suspension is centrifuged at 8000 rpm/20 min. The supernatant is acidified to pH=2, and immediately siderophore is extracted by adding equal volume of ethyl acetate in 50°C water bath .shaked to evaporate ethyl acetate layer, then the extract is placed in an oven at 50°C in open petri dishes to obtain dried extract. 6-Estimation of the dry weight of crude extract.

7-Determining the chemical nature of siderophore molecules; bacterial supernatant is used for assay by adding

1 ml of 2% of aqueous FeCl₃ to 1ml of sample. The result is positive by appearance of wine color absorbed at 490 nm in UV spectrophotometer [12].

Results and Discussion:

Identification Antimicrobial and **Susceptibility**

Nineteen isolates of A. baumannii from clinical sources; (5 from several sputum; 5 from wound swab; 4 from blood; 3 from urine and two isolates from tracheal secretion) are identified by growing of red colonies on CHRO Magar and depending on the identification results of Vitek 2 system (7). The results of the antimicrobial susceptibility are shown in Table(1).

Table . 1 Antimicrobial Susceptibility of A.baumannii Isolates

Antibiotic											
Isolate	PI	TI	CAZ	FEP	CRO	СТХ	MEM	TE	CIP	LEV	SXT
Ab1S	S	S	R	R	R	R	S	Ι	R	R	Ι
Ab2S	R	R	R	S	R	R	R	R	R	R	R
Ab3S	S	S	R	S	R	R	S	S	R	R	R
Ab4S	R	R	S	S	Ι	Ι	R	R	R	R	R
Ab5S	R	R	R	R	R	R	R	R	R	R	R
Ab6W	S	R	R	R	R	R	R	R	R	R	Ι
Ab7W	S	S	R	R	R	R	S	R	R	R	Ι
Ab8W	S	S	R	R	R	R	S	Ι	R	R	R
Ab9W	S	S	R	R	R	R	S	Ι	R	R	R
Ab10W	S	S	R	R	R	R	S	Ι	R	R	R
Ab11B	R	R	R	R	R	R	R	R	R	R	Ι
Ab12B	S	S	R	S	R	R	S	Ι	R	R	R
Ab13B	R	S	R	R	Ι	R	S	R	R	R	R
Ab14B	R	R	R	R	R	R	R	R	R	R	R
Ab15U	S	S	R	R	R	R	R	R	R	R	R
Ab16U	R	S	R	R	Ι	R	S	R	R	R	Ι
Ab17U	S	S	R	R	S	R	R	Ι	R	R	Ι
Ab18TS	R	R	R	R	R	R	R	R	R	R	R
Ab19TS	R	S	R	R	S	R	R	R	R	R	Ι

TI;Ticarcilin 75 µg S;Sensitive PI;Piperacilin 100µg R;Resistant I;Intermediate CAF;Ceftazidime 30 µg; FEP;Cefepime 30 µg CRO;Ceftriaxone 30 µg;CTX;Cefotaxime 30 µg; Meropenem 10 µg TE; Tetracycline 30 µg CIP; Ciprofloxacin 5 µg LEV; Levofloxacin MEM: SXT ; Trimethoprim-Sulfamethoxazole 1.25/23.75 µg 5 µg

Detection, Production and Extraction of Siderophores:

Investigation of chelating iron molecules (catecholate) secreted bv MDR-A.baumannii under iron restricted conditions has been carried out in this study. Five isolates (26%)are siderophore producers when tested on M9 medium as in Table (2):

Table.2	Sideror	ohore	Producing	Isolates
I anto	Diación	JIIUIC	IIVuuuung	Isolatos

Isolate	Ab1S	Ab2S		Ab3S	Ab4S	Ab55	6 Ab6W	Ab7W	Ab8W	Ab9W
Ab10W										
Туре	NP	NP	NP	NP	Р	NF	P NP	NP	NP	NP
Isolate	Ab11B	Ab12B		Ab13B	Ab1	4B	Ab15U	Ab16U	Ab17U	Ab18TS
Ab19TS										
Туре	Р	NP		NP	Р	NP	NP	Р	Р	NP
P:Produ	cer N	IP;Non p	rod	uce	S:spu	tum	W:wou	ınd E	B:blood	U:urine
TS: tracheal secretion										

S: tracheal secretion

The highlest producing isolate is Ab5S from sputum. Clear correlation is noticed between antimicrobial resistance and siderophore formation. Table (1) shows that the producer isolates are; Ab5S, Ab6W, Ab11B, Ab14B, Ab18TS with high resistance to antimicrobials. On M9 medium as mentiond earlier, the producer isolates colonies appear circular wrinkled and dried (Fig.1):



Fig.1 Colonies of Siderophore Producing Isolates on M9 Minimum Medium

Each colony in the Figure above represents the growth resulted from the stick touch, little and weak growth may be attributed to the poor and limited nutrients and depleted iron in M9 minimum medium.

Not all the isolates are siderophore producers and this agreed with the results of Yamamoto et al.,(1994) who report that 4 of 12 clinical A.baumannii strains examined are siderophore producers, indicative of strain -to-strain variation in the ability of acinetobactin production. [13] Sokol et al.,(1992) describe a novel siderophore from Pseudomonas cepacia (recently Burkholderia cepacia) cultures named azurechelin, and indicat that 88% of pathogenic isolates produced it. This compound correlates to bacterial virulence and may increase morbidity and mortality in patients of cystic fibrosis.[14] Bnyan al.,(2010) et showing that siderophore production is in 76.6% of uropathogenic Escherichia coli (UPEC) compared to 5% in E.coli fecal isolates thus siderophore production has been shown to be more frequent in E. coli from patients with UTI than in fecal isolates and is suggests that siderophore production positive isolates can be considered as UPEC. [15] Abass *et al.*,(2014) demonstrate

the role of two other genes in the virulence of UPEC; fimH (90.0)% and kpsMTII (72.0)% of *E.coli* isolated from UTI.

Other isolates have shown no growth on M9 minimum medium that may suggest variation in efficiency of siderophore production or they may form different siderophore other type of than catecholate according to Yamamoto et al. (1994) who detect and extract the acinetobactin (catecholate and hydroxamate functional groups) from A.baumannii.

The isolate Ab5S is inoculated in a specific minimum liquid medium and care is taken to use metal-free glass ware. Flasks and other glassware are kept in acid to remove all traces of metals from medium, inoculation have been done by sterile woody stick. The optimum conditions for maximum production occurs after 20hrs.at 30°C, pH 7, where no iron contamination was found. Previous studies indicate that the presence of iron can inhibit siderophore production as well as results indicating that iron-binding proteins, which may play a role in chelating the siderophorebound iron, are produced under ironstarved conditions [16,17]

Iron-binding proteins are present in membranes of cultures grown under iron limitation. Siderophores chelate iron and bacterial cell by outer supply to membrane receptors. iron is an important nutrient element for growth and maintenance, hence the siderophore molecules after 20 hrs. Become outside the bacterial cells -in the medium. The concentration of siderophores in the culture supernatant is maximal after 20 hrs. of growth which means that siderophore production occurrs in parallel with growth. Therefore extraction of these molecules should be bacterial done on filterate after precipitating of bacterial cells.

Because ethyl acetate layer is evaporated to dryness by shaking water –bath at 50° C, care should be taken from high temperatures which may denature the amino acids conjugated with the phenolates in catecholate siderophores.

Weight of crude extract is estimated which is equal to 115mg/l, it is considered low when compared to the result got by Hussien et al., (2013) where the weight of pyoverdin extract from P.aeruginosa is equal to 235 mg/l that may be attributed to the difference in the extraction medium and other experiment parameters as well as the producing microorganism. In another study [18], 200 mg/l also they the weight is extracted siderophores from P.aeruginosa .Chemical nature of the extracted molecules indicates catecholate (phenolates) structure because of wine colour of extract after adding 2% aq.FeCl₃ indicator absorbed at 490 nm in UV spectrophotometer.

Actis *et al.*, (1993) show in their work that different *A. baumannii* isolates are able to grow under iron-depleted conditions. The bacterial growth is accompanied by the formation of ironregulated catechol siderophores, independently of the bacterial plasmid content.

<u>Goel *et al.*</u>(1998) report that *A. baumannii* under iron restricted

conditions develop four high molecular outer membrane weight proteins (OMPs) of 88, 84, 80 and 77 kDa in iron depleted medium CDM-Fe which were absent in CDM + Fe medium, expressing iron regulated outer membrane proteins (IROMPs) along production of catechol with type siderophore is necessary to acquire iron from the external medium.

[19] Fukushima et al.,(2013) have documented that under iron starvation, siderophores are excreted, scavenge ferric ions and the complex is shuttled cell. inside the The microbial hydrophobicity decreases if Fe concentration is restricted which alters the surface protein receptors and leads to limitation of biofilm secretion [20].

[21] Pal and Gokarn (2010) have concluded that there is no significant difference occuring in the production of siderophore in commensal and clinical bacterial isolates. They suggested that siderophore production may be a necessary factor of virulence but not a determinant of virulence.

[22] Al- Muhanna et al., (2014) have discussed the correlation between siderophore and aerobactin gene. Isolates of K. pneumoniae that produce aerobactin are more virulent, but non siderophore producing isolates are less virulent. Also, they find that Κ. pneumoniae isolates totally produce siderophores are expressed aerobactin genes.

[23] **Naik** and Dubey (2011) document that low lead nitrates concentrations up to 0.5mM may enhance siderophore production in *P. aeruginosa*.

We conclude from our study that MDRbaumannii could produce Α. siderophores but in variable amounts among isolates. It is apparent that highly antimicrobial resistant isolates are siderophore producers.The extracted siderophore compound is from catecholate type.

References:

- Ali, S. S. and Vidhale, N. N. 2013. Bacterial Siderophore and their Application: A review Int. J. Curr. Microbiol. App. Sci. 2(12): 303-312.
- [2] Aguado- Santacruz, G. A. A.; Moreno-Gómez, B. A.; Jiménez-Francisco, B. B.; García-Moya E. B. and Preciado-Ortiz, R. E. 2012. Impact of the microbial siderophores and phytosiderophores on the iron assimilation by plants. Rev. Fitotec. Mex.; 35(7):9–21.
- [3] Chhibber, S.; Nag, D. and Bansal, S. 2013. Inhibiting biofilm formation by *Klebsiella pneumoniae* B5055 using an iron antagonizing molecule and a bacteriophage. BMC Microbiol.; 13(1):174–183.
- [4] Andrews, S. C.; Robinson, A. K. and Rodriguez-Quinones, F. 2003. Bacterial iron homeostasis. FEMS. Microbiol. Rev. 27(5):215-237.
- [5] Faraldo- Gómez, J. D. and Sansom, M. S. 2003. Acquisition of siderophores in Gram-negative bacteria. Nat. Rev. Mol. Cell Biol. 4(1): 105–116.
- [6] Mossialos, D. and Amoutzias, G. D. 2008. Role of siderophores in cystic fibrosis pathogenesis foes or friends. Int.J.Med.Microbiol.299 (15):87-98
- [7] Actis, L. A.; Tolmasky, M. E.; Crosa, L. M. and Crosa, H. J. 1993.
 Effect of Iron-Limiting Conditions on Growth of Clinical Isolates of *Acinetobacter baumannii*. J. Clin. Microbiol .10(2):2812-2815.
- [8] Yamamoto, S.; Okujo, N. and Sakakibara, Y. 1994. Isolation and structure elucidation of acinetobactin, a novel siderophore from *Acinetobacter baumannii*. Arch. Microbiol. 162(4):249-254.
- [9] Sambrook, J.; Fritsch, E. F. and Maniatis, T. 1989. Molecular cloning:a laboratory manual, 2nded. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N. Y. 29.

- [10] Shenker, M.; Oliver, I.; Heimann, M.; Hadar, Y. and Chen, Y. 1992. Modified M9 medium to detection siderophores of *Rhizobium ssp* in Tomato roots. J. Plant Nutr. 15(9):2173-2178.
- [11] Jadhav, R. S. and Desai, A. J. 1992. Isolation and characterization of siderophore from cowpea *Rhizobium* (peanut isolate). Curr. Microbiol. 24(3): 137-141.
- [12] Neilands, J.B. 1981. Microbial iron compounds. *Ann. Rev. Biochem.* 50(4):715-731.
- [13] Sokol, P. A.; Lewis, C. J. and Dennis, J. J. 1992. Isolation of a novel siderophore from *Pseudomonas cepacia*. J. Med. Microbiol. 36(11): 184-189.
- [14] Bnyan, I. H.; Bnyan H. A. and Ali, J. A. 2010. The Siderophore Production of *E. coli* Isolated from Urinary Tract Infection and Fecal Isolates. J. Babylon University/Pure and Applied Sciences/ 3(18):862-864.
- [15] Abass, Z. N.; Habib, K. A. and Abed, Z. A. 2014. Genotypic Study of Two Virulence Factors fimH and kps MTII in Uropathogenic *Escherichia coli* Isolates from Children Patients with Urinary Tract Infections. Bag. Sci. J. 11(4):1475-1480.
- [16] Goel, V. K.; Kapil , A.; Das, B. and Rao, D. N. 1998. Influence of iron on growth and extracellular products of *Acinetobacter baumannii*. Jap. J. Med. Sci. & Biol. 51(1):25-33.
- [17] Hussien, S. S.; Desouky, O. A.; Abdel-Haliem, M. E. F. and EL-Mougith, A. A. 2013. Enhancement the Production of Siderophores-Pyoverdine by *Pseudomonas aeruginosa* SHA 282 and Its Chelation with Thorium (IV). W. Res. J. Biotech. 1(1):17-23.
- [18] Schalk, I. 2008. Metal trafficking via siderophores in Gram-negative

bacteria: specificities and characteristics of the pyoverdine pathway. J. inorg. and Biochem., 102(10):1159-1169.

- [19] Fukushima, T.; Allred, B. E.; Sia, A. K.; Nichiporuk, R.; Andersen, U. N. and Raymond, K. N. 2013. Grampositive siderophore-shuttle with iron-exchange from Fe-siderophore to apo-siderophore by *Bacillus cereus YxeB*. Proc. Natl. Acad. Sci. U.S.A. 110(30): 13821–13826.
- [20] Simões, L. C.; Simões, M. and Vieira, M. J. 2007. Biofilm interactions between distinct bacterial genera isolated from drinking water. Appl. Environ. Microbiol.; 73(8) :6192–6200.

- [21] Pal, R. B. and Gokarn, K. 2010. Siderophores and Pathogenecity of Microorganisms. J. Biosci. Tech.1 (3).127-134.
- [22] Al- Muhanna, A. S.; Al-Rediany, R. S. and Alzuhairi, M. A. 2014. Molecular characterization of *aerobactin* gene among *Klebsiella* isolated from Wound and Burn Infections Int. J. Curr. Microbiol. App. Sci. 3(5): 26-31.
- [23] Naik, M. M. and Dubey, S. K. 2011. Lead-enhanced siderophore production and alteration in cell morphology in a Pb-resistant *Pseudomonas aeruginosa* strain 4EA. Curr. Microbiol.62(2):409-14.

إنتاج وإستخلاص مركبات السايدروفور ـ نوع الكاتيكولات ـ من بكتريا Acinetobacter baumannii متعددة المقاومة للمضادات الحيوية

قسم علوم الحياة، كلية العلوم، الجامعة المستنصرية، بغداد، العراق.

الخلاصة:

السايدروفورات (المركبات الحاملة أو الناقلة للحديد) هي جزيئات عضوية ذات أوزان جزيئية واطئة تنتجها الأحياء المجهرية عند نموها في ظروف يكون فيها عنصر الحديد قليل أو معدوم تم في الدراسة الحالية التحري عن مركبات السايدروفور وظروف انتاجها واستخلاصها من عزلات سريرية لبكتريا A.baumannii متعددة المقاومة للمضادات الحياتية. جمعت 120 عزلة لبكتريا سالبة لصبغة غرام غير مخمرة لسكر اللاكتوز من ثلاث مستشفيات في مدينة بغداد ولمدة ثلاثة شهور شخصت العزلات مبدئيا بالاختبارات التشخيصية القياسية: الاختبارات الكيموحيوية واشرطة (API 20 NE) وظهر أن 19 عزلة منها تعود أبكتريا

متعددة المقاومة للمضادات الحيوية والتي تم تأكيد تشخيصها بأستخدام الوسط الملون Vitek 2 الخاص بالاسينيتو فضلا عن نظام Vitek 2 . اخضعت جميع العزلات لفحص الحساسية CHROMagar الخاص بالاسينيتو فضلا عن نظام Vitek 2 . اخضعت جميع العزلات لفحص الحساسية للمضادات الحياتية . تم التحري عن العزلات المنتجة للمركبات الناقلة للحديد بتنميتها في الوسط المتدني M9 المضادات الحياتية . تم التحري عن العزلات المنتجة للمركبات الناقلة للحديد بتنميتها في الوسط المتدني M9 المضادات الحياتية . تم التحري عن العزلات المنتجة للمركبات الناقلة للحديد بتنميتها في الوسط المتدني M9 المضادات الحياتية . تم التحري عن العزلات المنتجة للمركبات الناقلة للحديد بتنميتها في الوسط المتدني M9 المضادات الحياتية . تم التحري عن العزلات المنتجة للمركبات الناقلة للحديد بتنميتها في الوسط المتدني M9 المضادات الحياتية . م التحري عن العزلات المنتجة للمركبات الناقلة للحديد بتنميتها في الوسط المتدني M9 المضادات الحياتية . م التحري عن العزلات المنتجة للمركبات الناقلة للحديد بتنميتها في الوسط المتدني M9 المضادات الحياتية . م التحري عن العزلات المنتجة للمركبات الناقلة للحديد بتنميتها في الوسط المتدني M9 المضاد الحياتية . م التحري عن أن 26% منها كانت منتجة لجزيئات السايدروفور أذ كانت العزلة الأوفر إنتاجاً هي مع ملحدها هو القشع (Sputu) . نميت هذه العزلة في وسط تركيبي خاص لأنتاج الجزيئات الناقلة للحديد وأستخلصت هذه الجزيئات بواسطة مركب خلات الإيثيل وتم تقدير وزن المستخلص الجزيئات الناقلة للحديد وأستخلوست هذه الجزيئات الماليمينيات والتي ظهر أنها من الكاتيكولات (فينولات).

الكلمات المفتاحية: A.baumannii، السايدروفورات، الكاتيكولات