

## Synthesis and Characterization of New Selenonitron Derivative and Its Effect on Breast Cancer Cell Line Viability *in Vitro*

Batool S. Haddad<sup>1\*</sup>

Kawthar K. Hassan<sup>2</sup>

Received 15/7/2018, Accepted 14/3/2019, Published 22/9/2019



This work is licensed under a [Creative Commons Attribution 4.0 International License](https://creativecommons.org/licenses/by/4.0/).

### Abstract:

New nitron and selenonitron compounds were synthesized. The condensation method between N-(2-hydroxyethyl) hydroxylamine and substituted carbonyl compounds such as [benzil, 4, 4'-dichlorobenzil and 2,2'-dinitrobenzil] afforded a variety of new nitron compounds while the condensation between N-benzylhydroxylamine and substituted selenocarbonyl compounds such as [di(4-fluorobenzoyl) diselenide and (4-chlorobenzoyl selenonitrile)] obtained selenonitron compounds. The condensation of N-4-chlorophenylhydroxylamine with dibenzoyl diselenide obtained another type of selenonitron compounds. The structures of the synthesized compounds were assigned based on spectroscopic data (FT-IR, <sup>1</sup>H-NMR, <sup>13</sup>C-NMR, MS and elemental analysis. The result spectra and the result of elemental analysis were verified the expected structure. The results of new selenonitron derivative (N<sub>1</sub>) effect on the MDA-MB-231 breast cancer cells *in vitro* revealed that is an increase in the proliferation of cells with increased concentrations of selenonitron at all periods of the times.

**Key words:** Cell viability, MDA-MB-231 breast cancer cell line, Nitrones, Proliferation, Selenocarbonyl, Selenonitrones.

### Introduction:

Nitron has been extensively investigated, because of their utility as versatile synthetic intermediates. Nitrons are very important compounds due to their successful application, they can be used as regulators and modifiers regulators of molecular weight in radical polymerization building blocks (1). The scientists also found that nitron can be used as building blocks in the synthesis of various natural and biologically active compounds (2).

For the preparation of nitrones, the most popular method is the condensation of aldehydes or ketones with N-monosubstituted hydroxylamines (3). However, this method is difficult to be applied to the preparation of ketonitrones having bulky groups. The synthesis of N-monosubstituted hydroxylamine is being carried out by the reaction of (benzylchloride and 2-hydroxyethylchloride) with hydroxylamine hydrochloride in presence of sodium carbonate (4). Organoselenium compounds have been aimed as targets of interest in organic synthesis.

Dibenzoyl diselenide [(PhCOSe)<sub>2</sub>] is an important example of this class showing several pharmacological properties (5-7). The synthesis of dibenzoyl diselenide is being carried out by the alkaline hydrolysis of substituted benzoylselenonitrile (the synthesis of benzoylselenonitrile is being carried out by the reaction between benzoyl chloride and potassium selenonitrile) (8-10). Benzil (PhCO)<sub>2</sub> were prepared via the condensation reaction of substituted benzaldehyde. Thus, condensation of various substituted benzil with N-(2-hydroxyethyl) amine makes it possible to synthesize a variety of nitron compounds (11), while the condensation of various substituted dibenzoyl diselenide and benzoyl selenonitrile with N-benzylhydroxylamine and N-phenylhydroxylamine makes it possible to synthesize a variety of selenonitron compounds (12,13). The condensation is carried out under mild conditions without affecting functional groups. Nitrones due to their photochemistry reactivity as well as their excellent radical spin-trapping capability leading to their use as antioxidants because of stable nitroxyl radicals. Organoselenium compounds also, such as containing di aryl diselenide group are exhibited significant antimicrobial (14-17), cytotoxic, antidepressant, antileukemic, antibiotic and antiurease activities

<sup>1</sup> Department of Chemistry, College of Science, University of Basrah, Basrah, Iraq

<sup>2</sup> Department of Human Anatomy, College of Medicine, University of Basrah, Basrah, Iraq

\*Corresponding author: [kawther.hassan@uobasrah.edu.iq](mailto:kawther.hassan@uobasrah.edu.iq)

(18-20). Selenium is an essential trace element for humans and many other forms of life (21). It is an extremely vital mineral for the human body as it increases immunity (22). Also reducing inflammation and regulating blood pressure (23). According to Institute of Medicine, Food and Nutrition Board (24) the current recommended daily allowance for selenium is 55 µg/ day for a healthy adult. The toxic levels are estimated to occur with intakes of the order of 350–700 µg/day (25). Deficiency of selenium is associated with several disease conditions such as immune impairment (26). According to Rayman (27) study, consuming plenty of naturally occurring selenium has positive antiviral effects, essential for successful male and female fertility and reproduction, and also reduces the risk of autoimmune and thyroid diseases. Selenium plays a role in cancer prevention and treatment (28). On other hands selenium-induced cell migration and proliferation in some cases (29-30).

The aim of the study is to synthesis five compounds of selenonitrene and studies the effect of less toxic compound ( $N_1$ ) on viability MDA-MB-231 breast cancer cell line *in vitro*.

### Materials and Methods:

All chemicals were obtained from standard commercial sources. The synthesized compounds were prepared according to literature procedures. Carbonyl selenonitrile compounds have been carried out by the reaction of the corresponding benzoyl chloride with potassium selenocyanate. The synthesis of dibenzoyl diselenide is being carried out by the alkaline hydrolysis of substituted carbonylselenonitrile. Substituted benzil compounds were synthesized by the condensation reaction of substituted benzaldehyde. Selenonitrene compounds were synthesized by the condensation of dibenzoyl diselenide and selenonitrile with [N-benzyl hydroxyl amine and N-phenyl hydroxyl amine]. Nitrene compounds were synthesized by the condensation of benzil with N-(2-hydroxyethylhydroxylamine).

Melting points were measured with an Electrothermal IA 9200 apparatus. FT-IR spectra were recorded using a Bruker spectrometer.  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra were recorded in DMSO and  $\delta$  units downfield from internal reference  $\text{Me}_4\text{Si}$  using a 300 MHz Bruker NMR spectrometer. The mass spectra were recorded on a Shimadzu QP GC-MS instrument. Elemental analysis was carried out on a Vario EL III instrument (31-36).

### General Procedure for the Preparation of Carbonylselenonitrile (31, 32):

A solution of KCN (0.8gm) in absolute ethanol (50 ml) was added to a solution of selenium metal

(1 gm.; 125 mole) in absolute ethanol (15 mL). The reaction mixture was refluxed for 3 h and then a solution of substituted benzoyl chloride (206 ml 0.025 mole) was added and refluxed for 2 h. The precipitate was filtered and then recrystallized from ethanol.

### General Procedure for the Preparation of Dibenzoyl diselenide (33):

A solution of carbonylselenonitrile (0.28 gm; 1.02 mole) in absolute ethanol (25 ml) was added to a solution of sodium hydroxide (0.16 gm; 2.47 mole) in absolute ethanol (15 mL). The reaction mixture was stirred at 40 ° C for 50 min and then the mixture was refluxed for 1 h. A solution of substituted benzoyl chloride (206 ml 0.025 mole) was added and refluxed for 2 h. The reaction mixture was cooled to R.T. and filtered. The filtrate was acidified with 10% HCl. A red solid compound was obtained, washed with a small amount of benzene and dried. The precipitate recrystallized from a mixture of methanol and dichloro methane (1:4), gave a red solid compound.

### General Procedure of Preparation of Benzil (34):

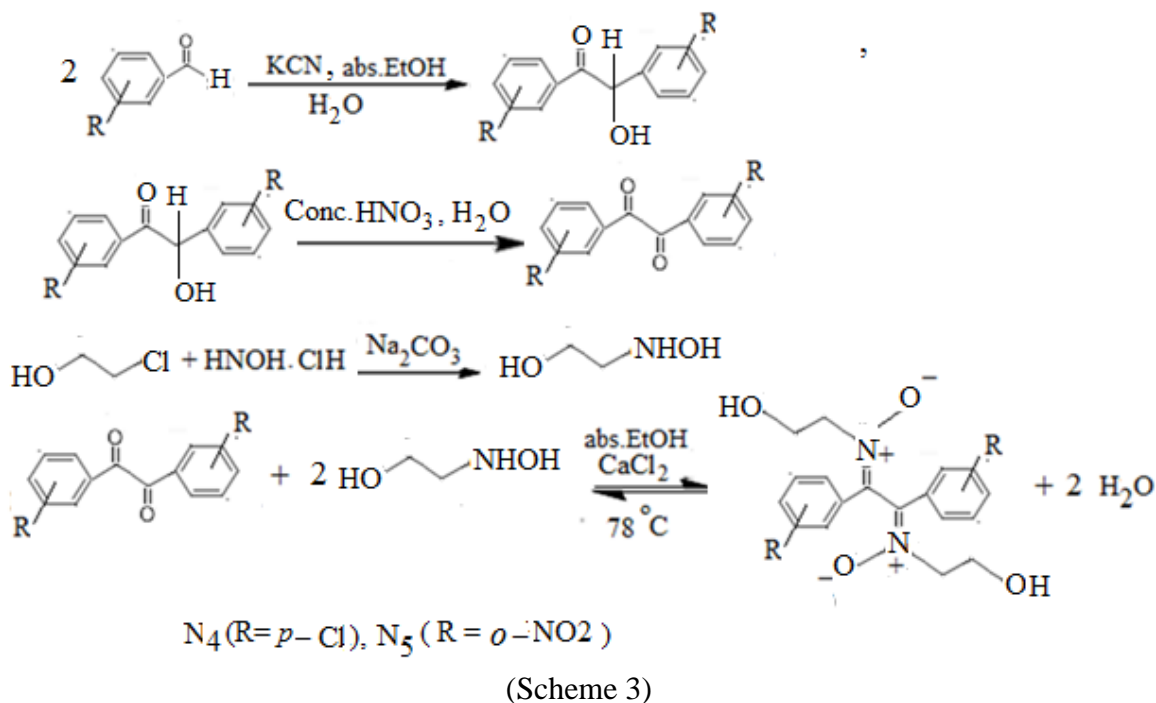
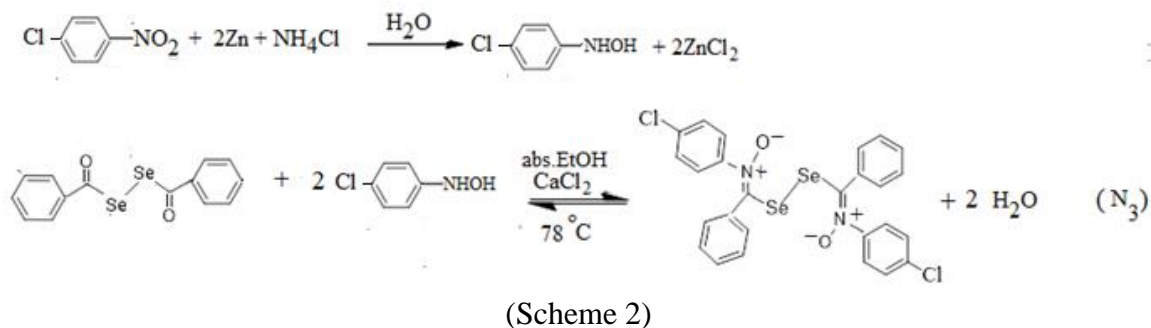
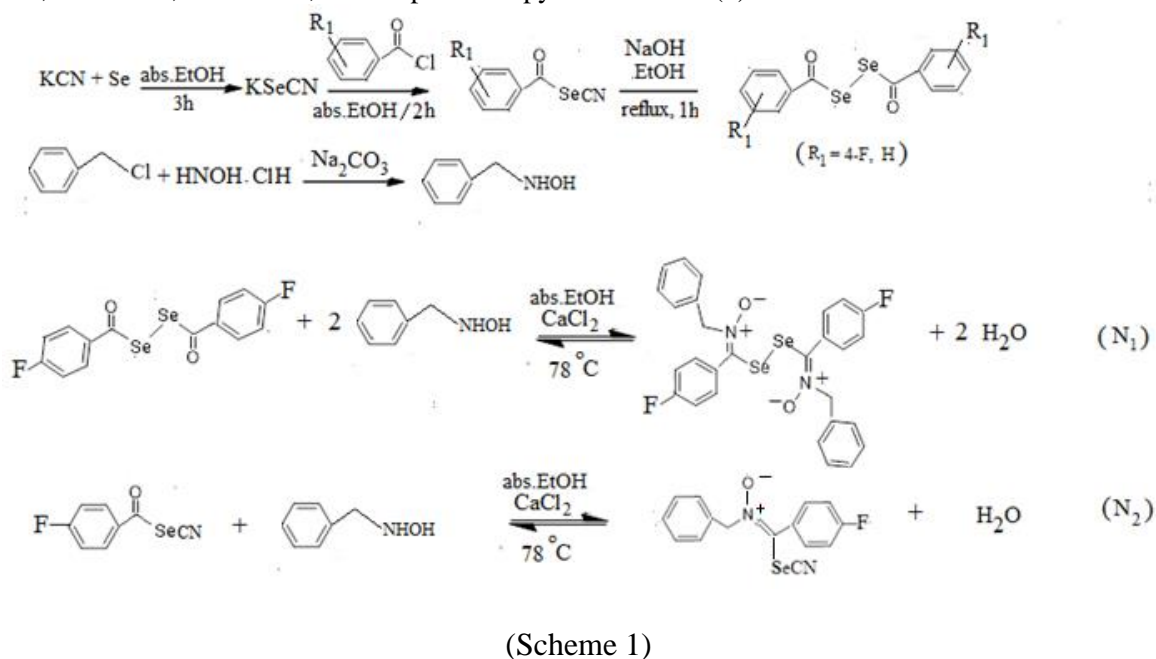
A solution of KCN (2.0 gm.; 0.036 mol) in  $\text{H}_2\text{O}$  (15 ml) was added to a solution of benzaldehyde (3.7 gm.; 0.03 mole) in ethanol (30 ml). The reaction mixture was refluxed for 0.5 h. The reaction mixture was cooled. A yellow solid compound of benzoin was obtained, washed with (1: 1- water: ethanol) and dried. (14 ml) of conc. nitric acid was added to a solution of benzoin (0.4 gm.; 0.018 mol). The reaction mixture was stirred and heated at 50 ° C for 11 min and then (75 ml) of  $\text{H}_2\text{O}$  was added. The reaction mixture was cooled and a yellow solid compound of benzil was obtained. The mixture was filtered and the precipitate recrystallized from methanol.

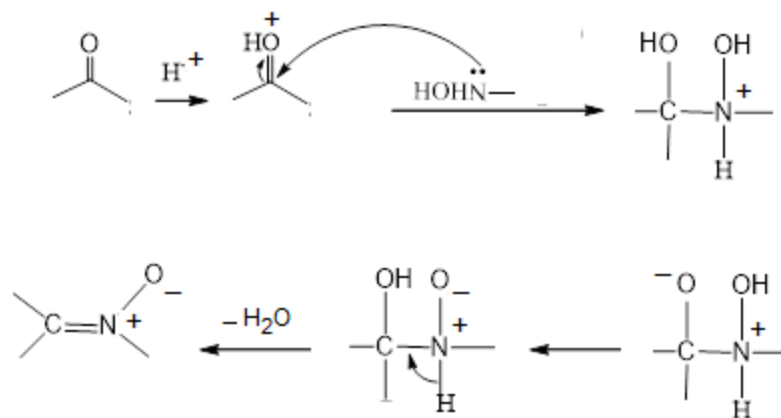
### General Procedure of Synthesis Nitrene (35, 36):

A solution of diphenyl diselenide for synthesis ( $N_1$ ,  $N_3$ ), benzoylselenonitril for synthesis  $N_2$  and benzil for synthesis ( $N_4$  and  $N_5$ ), (0.02 mole) in absolute ethanol (30 ml) was added to a solution of anhydrous  $\text{CaCl}_2$  (15 gm) in absolute ethanol (30 ml) placed in a (250 ml) one-necked round-bottomed flask. The reaction mixture was stirred at 50 ° C for 30 min and then a solution of N-benzylhydroxylamine [for synthesis ( $N_1$  and  $N_2$ ) as in (scheme 1)], (N-4-chlorophenyl hydroxylamine) [for synthesis ( $N_3$ ) as in (scheme 2)], N-(2-hydroxyethylamine) [for synthesis ( $N_4$  and  $N_5$ ) as in (scheme 3)], (0.02 mole) in absolute ethanol 30 ml was added. 3 drops of benzene sulfonic acid were added and the mixture of ( $N_1$ ,  $N_2$ ,  $N_4$  and  $N_5$ ) was refluxed in the dark overnight. While the mixture reaction of ( $N_3$ ) was kept with stirring in dark, overnight at room temperature. The reaction mixture was cooled and the precipitate was filtered

and then recrystallized from absolute ethanol. The mechanism of reaction explains in the (scheme 4). All the synthesized compounds were characterized by FTIR,  $^1\text{H}$  NMR,  $^{13}\text{C}$  NMR, mass spectroscopy

and Elemental analysis (CHN) and all the results were in agreement with the theoretical calculations. Physical properties of nitrones are represented in the table (1).





(Scheme 4)

The mechanism of the formation of nitron compounds

**Table 1. Physical properties of nitrones**

Compounds	M. p °C	Time of reaction	Yield (%)
N <sub>1</sub>	150-152	33	63
N <sub>2</sub>	161-163	20	60
N <sub>3</sub>	101-103	11	69
N <sub>4</sub>	90-92	17	77
N <sub>5</sub>	123-125	14	71

#### Cell culture:

The MDA-MB-231 breast cancer cell line (Beatrice Hunter Cancer Research Institute, Canada) is an epithelial, human breast cancer cell line that was established from a pleural effusion of a 51-year-old Caucasian female with a metastatic mammary adenocarcinoma and is one of the most commonly used breast cancer cell lines in medical research laboratories (37).

Cell lines were grown, maintained and passaged in a humidified incubator at 37°C with 5% CO<sub>2</sub> atmosphere. Cells were cultured as a monolayer in high glucose DMEM (Dulbecco's modified Eagle's medium) supplemented with 10% fetal bovine serum (FBS) (both from Invitrogen), and antibiotics (100 IU/mL penicillin and 100 μL/mL streptomycin) according to the protocol of Beatrice Hunter Cancer Research Institute, Canada.

#### Cell viability assay

According to the protocol of Beatrice Hunter Cancer Research Institute, Canada, The anti-proliferative effects of selenonitron on MDA-MB-231 cells were examined using MTT assay (38). The Cell lines were seeded in 96-multiwell plates at a density of 4500 cells per well for 24h. before exposure to the different concentrations of compounds for 24, and 48 h respectively. Fifty μM of MTT was added to each well after the medium was decanted. Followed by 4 h. incubation. The MTT solution was discarded and the purple formazan crystals formed by the mitochondrial dehydrogenase enzymes were dissolved in 0.1N HCl in isopropanol. The absorbance was read on the

Synergy Power wave plate reader at 570 nm. The percent of MTT compared to the vehicle control cells was calculated at viability.

Data were shown as a mean ± standard error and were analyzed using one-way analysis of variance (ANOVA) according to Statistical Package for Social Science (SPSS). The level of significance was set at α = 0.05, and statistically significant changes were indicated in figures as follows: \*P<0.05, \*\*P<0.01, \*\*\*P<0.001, compared to control.

#### Results and Discussion:

Treatment of N- hydroxylamine with selenocarbonyl derivatives gave selenonitron compounds N<sub>1</sub>, N<sub>2</sub> as shown in (Scheme 1). The reaction of N- hydroxylamine with carbonyl derivatives gave nitron compounds N<sub>4</sub> and N<sub>5</sub>, as shown in (Scheme 3) in good yields (77-71)%. The yield of selenonitrones N<sub>1</sub>, N<sub>2</sub> and N<sub>3</sub> (63, 60, 69) % This is mainly due to the easily decomposed of organoselenium compounds in light and moisture. All the IR spectra of nitrones showed the disappearance of (C=O) of carbonyl band in the region (1700-1670) cm<sup>-1</sup> and the appearance peaks at the range (1619-1604) cm<sup>-1</sup> due to (C=N) stretching, also peaks at the range (1290-1280) cm<sup>-1</sup> due to (N-O) stretching. The (C=C) stretching showed peaks at the range (1585-1568) cm<sup>-1</sup> and the (C-H) stretching aromatic ring showed peaks at the range (3209-3000) cm<sup>-1</sup>, while showed peaks at the range (2890-2840) cm<sup>-1</sup> due to (C-H) aliphatic. The (C-Se) stretching of N<sub>1</sub>, N<sub>2</sub> and N<sub>3</sub> were showed in the range (571, 584 and 583) cm<sup>-1</sup>. The (CN) stretching of nitron (N<sub>3</sub>) was shown in the range (2100) cm<sup>-1</sup>. All the <sup>1</sup>H NMR spectra of selenonitrones and nitrones were showed multiplet signals at the range δ= (7.78-7.60) ppm for the aromatic protons rings. The spectra of nitrones were showed signals in the region δ=4.06 ppm and

$\delta=4.50$  ppm for the two groups of  $\text{CH}_2$  and showed signals in the region  $\delta= (3.6-3.2)$  ppm (OH) stretching. Mass spectroscopy gives the molecular ion and other fragments which indicated the structure of synthesized compounds. All the nitron compounds have been diagnosed by elemental analysis, and all the results were in agreement with the calculated values.

**(N<sub>1</sub>) N,N'-(diselanediyil bis ((4-fluoro phenyl) methan-1-yl-1-ylidene))bis (1-phenylmethanamine oxide)**

Yellow crystals; yields: 60%; m.p. 150-152 °C; IR (v cm<sup>-1</sup>): 1619 (v C=N), 1568 (v C=C), 1286 (v N-O), 571 (v C-Se), 1170 (v C-F), 3209 (v CH aromatic), 2885-2876 (v CH aliphatic); <sup>1</sup>H NMR (DMSO, 300 MHz;  $\delta$  ppm)  $\delta$ : 7.84 (m, 4H, ph), 7.69 (m, 5H, Ph), 4.26 (s, 2H, CH<sub>2</sub>,  $J = 7$  Hz); <sup>13</sup>C NMR (DMSO, 300 MHz;  $\delta$  ppm): 57.04, 126.10, 128.23, 129.14, 131.65, 132.66, 140.27, 142.50, 145.71, 162.33; Mass spectrum: m/z (relative intensity): 614(M<sup>+</sup>). Anal. calcd. for C<sub>28</sub>H<sub>22</sub>N<sub>2</sub>O<sub>2</sub>F<sub>2</sub>Se<sub>2</sub>: C 54.72, H 3.58, N 4.56. found: C 54.61, H 3.60, N 4.37.

**(N<sub>2</sub>) 1-Phenyl-N-(1,1-(4-fluorophenyl) selenonitrile methylene) methanamine oxide**

Yellow crystals; yields: 71%; m.p. 101-102 °C; IR (v cm<sup>-1</sup>): 1612 (v C=N), 1585 (v C=C), 1290 (v N-O), 584 (v C-Se), 766 (v C-Cl), 2100 (v CN), 3112 (v CH aromatic), 2889-2877 (v CH aliphatic); <sup>1</sup>H NMR (DMSO, 300 MHz;  $\delta$  ppm)  $\delta$ : 7.66 (m, 5H, ph), 7.10 (m, 4H, ph), 4.06 (t, 2H, CH<sub>2</sub>,  $J = 7.1$  Hz); <sup>13</sup>C NMR (DMSO, 300 MHz;  $\delta$  ppm)  $\delta$ : 35.83, 114.62, 123.97, 124.68, 126.69, 128.34, 129.44, 130.41, 131.58, 133.82, 162.11; Mass spectrum: m/z (relative intensity): 333(M<sup>+</sup>). Anal. calcd. for C<sub>15</sub>H<sub>11</sub>N<sub>2</sub>OFS<sub>2</sub>: C 54.05, H 3.30, N 8.40. found: C 54.12, H 3.24, N 8.51.

**(N<sub>3</sub>) N,N'-(diselanediyilbis (phenyl methan-1-yl-1-ylidene))bis(4-chloroaniline oxide).**

Orange crystals; yields: 63%; m.p. 163-164 °C; IR (v cm<sup>-1</sup>): 1614 (v C=N), 1578 (v C=C), 1280 (v N-O), 583 (v C-Se), 760 (v C-Cl), 3111 (v CH aromatic); <sup>1</sup>H NMR (DMSO, 300 MHz;  $\delta$  ppm)  $\delta$ : 7.54 (m, 4H, Ph), 7.90 (m, 5H, Ph), 4.12 (s, 2H, CH<sub>2</sub>,  $J = 6.8$  Hz); ; <sup>13</sup>C NMR (DMSO, 300 MHz;  $\delta$  ppm): 55.69, 128.09, 129.15, 130.15, 131.38, 141.50, 143.21, 144.55, 161.43; Mass spectrum: m/z (relative intensity): 646(M<sup>+</sup>). Anal. calcd. for C<sub>28</sub>H<sub>22</sub>N<sub>2</sub>O<sub>2</sub>Cl<sub>2</sub>Se<sub>2</sub>: C 52.01, H 3.40, N 4.33. found: C 52.22, H 3.29, N 4.40.

**(N<sub>4</sub>) N,N'-(bis (4-chlorophenyl methan-1-yl-1-ylidene)) di(2-hydroxyethanamine oxide).**

Yellow crystals; yields: 77%; m.p. 127-129 °C; IR (v cm<sup>-1</sup>): 1604 (v C=N), 1581 (v C=C), 1285 (v N-O), 795 (v C-Cl), 3110 (v CH aromatic), 2890-2865 (v CH aliphatic), 3310 (v OH);; <sup>1</sup>H NMR

(DMSO, 300 MHz;  $\delta$  ppm)  $\delta$ : 7.70 (m, 4H, ph), 4.13 (t, 2H, CH<sub>2</sub>,  $J = 6.9$  Hz), 4.50 (t, 2H, CH<sub>2</sub>,  $J = 7.1$  Hz), 3.2 (s, OH); <sup>13</sup>C NMR (DMSO, 300 MHz;  $\delta$  ppm): 123.11, 124.08, 127.44, 129.17, 130.00, 132.01, 160.28; <sup>9</sup>Mass spectrum: m/z (relative intensity): 396(M<sup>+</sup>). Anal. calcd. for C<sub>18</sub>H<sub>18</sub>N<sub>2</sub>O<sub>4</sub>Cl<sub>2</sub>: C 54.54, H 4.54, N 7.07. found: C 54.61, H 4.40, N 7.20.

**(N<sub>5</sub>) N,N'-(bis (2-nitrophenyl methan-1-yl-1-ylidene)) di(2-hydroxyethanamine oxide).**

Orange crystals; yields: 69%; m.p. 150-151 °C; IR (v cm<sup>-1</sup>): 1615 (v C=N), 1580 (v C=C), 1283 (v N-O), (s) 1390 - (as) 1520 (v NO<sub>2</sub>), 3118 (v CH aromatic), 2889-2858 (v CH aliphatic), 3308 (v OH);; <sup>1</sup>H NMR (DMSO, 300 MHz;  $\delta$  ppm)  $\delta$ : 7.60 (m, 4H, ph), 4.09 (2H, CH<sub>2</sub>), 4.33 (2H, CH<sub>2</sub>), 3.01 (s, OH); <sup>13</sup>C NMR (DMSO, 300 MHz;  $\delta$  ppm): 121.34, 125.10, 128.22, 129.15, 131.55, 132.67, 160.23; <sup>9</sup>Mass spectrum: m/z (relative intensity): 418(M<sup>+</sup>). Anal. calcd. for C<sub>18</sub>H<sub>18</sub>N<sub>4</sub>O<sub>8</sub>: C 51.67, H 4.30, N 13.39. found: C 51.42, H 4.11, N 13.43.

A comparison was made between the five compounds prepared by a toxicity assay and found that the N<sub>1</sub> containing selenium was less toxic than the other four compounds therefore, this compound was tested only.

**Cytotoxic effects of selenonitrone (N<sub>1</sub>) on MDA-MB-231 cancer cell line *in vitro* .**

The result of the present study an effect of selenonitrone on proliferation of MDA-MB-231 cancer cell line has been shown in the table (2). It was highly significant ( $P \leq 0.001$ ) between cells treated with selenonitrone and treated with control (DMSO). The study also showed highly significant difference among all concentrations of selenium compound at all periods of the treatment. Highly significant differences ( $P \leq 0.001$ ) were found in the interaction between compounds and concentrations after 24 and 48 h.

**Table 2. Analysis of variance for effect of different compound, concentrations of selenonitrone and relation between (compounds and concentrations) on the proliferation of MDA-MB-231 breast cancer cells after 24 and 48 hours.**

S.O.V.	MS.	
	24h.	48h.
Compounds (Selenonitrone and DMSO)	0.021855***	0.183612***
Concentrations	0.043511***	0.096764 ***
Concentrations & Compounds	0.076179 ***	0.029043 ***

S.O.V.= source of variance , MS=mean square, \*\*\*=( $P \leq 0.001$ )

In the table (3) showed at P-value less than 0.5 indicates, that there significant different effect

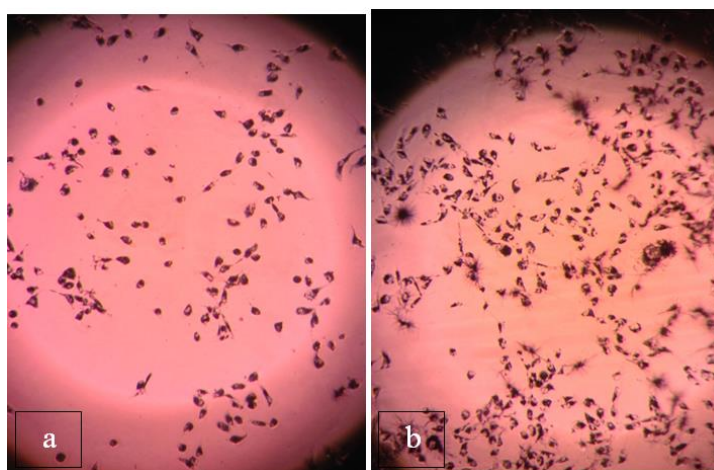


between cell treated with selenonitron and DMSO vehicle control. The toxicity effect of selenonitron varied with different concentrations. In all concentrations from 0.7  $\mu\text{g/ml}$  and up to 500  $\mu\text{g/ml}$  after 24 h. had shown a significant effect on MDA-MB-231 breast cancer cell line as compared to the vehicle control group (DMSO), the viability of the

cell (O.D optical density) was increased in all concentrations ascending from  $0.436 \pm 0.0354$  to  $0.696 \pm 0.0849$ . In (Fig.1) shown that decrease of toxicity effect of selenonitron ( $N_1$ ) on MAD-MB breast cancer cells proliferation as a compared with control group.

**Table 3. Mean and SD for effect of different concentrations of selenonitron ( $N_1$ ) and DMSO on the proliferation of MDA-MB-231 breast cancer cells after 24 hours.**

Compound	Selenonitron ( $N_1$ )	DMSO	Media	P value ( $p \leq 0.5$ ) between comp. & DMSO
Conc. ( $\mu\text{M}$ )	Absorbance (viability) and std.	Absorbance (viability) and std.	Absorbance (viability) and std.	0.0001992 ***
0.7	$0.436 \pm 0.0354$	$0.442 \pm 0.0385$	$0.519 \pm 0.0286$	
5	$0.436 \pm 0.0354$	$0.444 \pm 0.0315$		
25	$0.463 \pm 0.0375$	$0.461 \pm 0.0167$		
50	$0.466 \pm 0.0212$	$0.480 \pm 0.0012$		
250	$0.583 \pm 0.0615$	$0.426 \pm 0.0255$		
500	$0.696 \pm 0.0849$	$0.422 \pm 0.0339$		



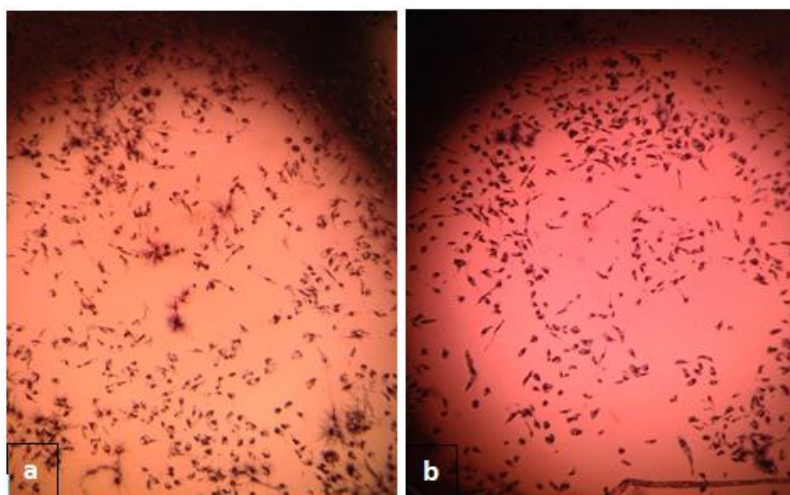
**Figure 1. MAD-MB breast cancer cells proliferation incubated for 24 h. (a) treated with DMSO. (b) treated with selenonitron ( $N_1$ ) at 500  $\mu\text{M}$  concentration.**

After 48 h. treatment, table (4) showed that there significant effect ( $P \leq 0.5$ ) between selenium compound and DMSO vehicle control. Also showed higher increased of a stimulatory effect of selenonitron ( $N_1$ ) on MAD-MB breast cancer cells proliferation at all concentrations more than incubation after 24h. The effect of selenonitron was increased according to the increase of the

concentration as compared to control group  $0.502 \pm 0.0625$ ,  $0.520 \pm 0.0292$ ,  $0.552 \pm 0.00820$ ,  $0.571 \pm 0.0340$ ,  $0.624 \pm 0.0341$  and  $0.747 \pm 0.0459$  respectively. Microscopic observations of selenium-treated cells revealed an active proliferation of malignant cells at 500  $\mu\text{M}$  concentration compared with untreated control cells (Fig. 2).

**Table 4. Mean and SD for effect of different concentrations of selenonitron ( $N_1$ ) compound and DMSO on the proliferation of MDA-MB-231 breast cancer cells after 48 hours.**

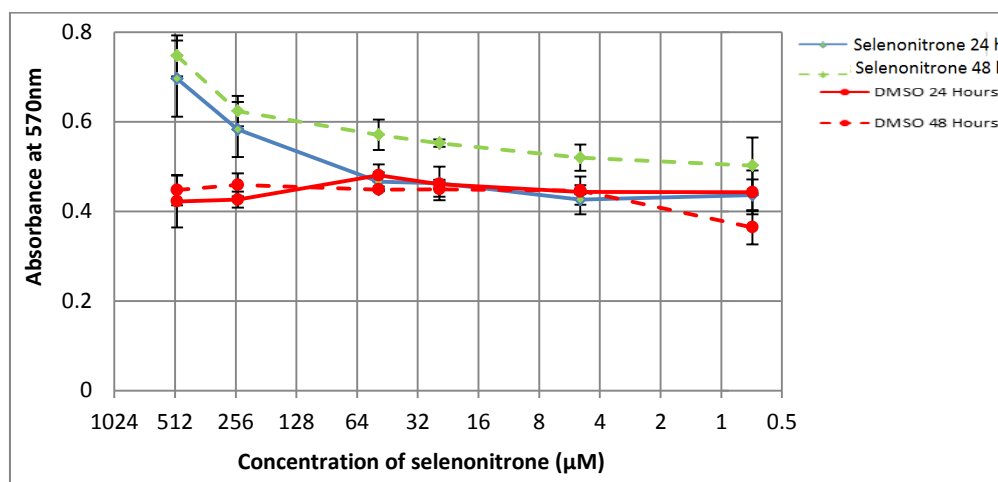
Compound	Selenonitron ( $N_1$ )	DMSO	Media	P value ( $\leq 0.5$ ) between comp. & DMSO
Conc. ( $\mu\text{M}$ )	Absorbance (viability) and std.	Absorbance (viability) & std.	Absorbance (viability) and std.	0.0001833 ***
0.7	$0.502 \pm 0.0625$	$0.364 \pm 0.0488$	$0.431 \pm 0.056$	
5	$0.520 \pm 0.0292$	$0.446 \pm 0.0056$		
25	$0.552 \pm 0.0082$	$0.449 \pm 0.0094$		
50	$0.571 \pm 0.0340$	$0.448 \pm 0.0247$		
250	$0.624 \pm 0.0341$	$0.459 \pm 0.0180$		
500	$0.747 \pm 0.0459$	$0.448 \pm 0.0581$		



**Figure 2. MAD-MB breast cancer cells proliferation incubated for 48.(a) treated with DMSO. (b)treated with selenonitron at 500 µM concentration**

Figure 3 demonstrated that selenonitron was more effect after 48 h. than 24 h. on the growth of MAD-MB breast cancer cells in comparison to DMSO

control groups. This effective increased in time and concentration- dependent manner.



**Figure 3. Absorbance of MAD-MB breast cancer cells treated with selenonitron ( $N_1$ ) and with DMSO concentrations after incubation for 24 and 48 hours**

The goal of current study is to understand the roles of the selenium in cancer prevention but it is revealed unexpected results on MDA-MB-231 breast cancer cell lines, although the experiment was repeated twice and gave the same results. The results indicated that selenium applied for 24 h and 48 h. stimulated cell proliferation as a compared to untreated control cells in a concentration and time-dependent manner, that is compatible with a previous study carried by Scott *et al.*(39) found the selenium did not prevent cancer .

The data in study of Grana & Reddy (40) and Kim, *et al.* (41) support the result that obtained in the present study, they found that the selenium at low concentration leads to the promotion of cell cycle progression due to up-regulates multiple key cell cycle-related gene mRNA levels or by enhances total cellular phosphorylated proteins in

HL-60 cells in serum-free culture media , particularly the G2/M transition, and/or a reduction of apoptosis primarily in G1 cells (42) ,while the Shin-Hyung *et al.* (35) showed the selenium treatment enhanced the cell growth, with 15 ng /ml of selenium the most effective in stimulating cell proliferation among various concentrations of selenium tested.

The current study agrees with a study of Petra *et al.* (43) have been reported that selenoprotein has cancer-promoting effects and agrees with Shin-Hyung (30) that demonstrated the selenium stimulates the cell growth and the proliferation of 3T3-L1 preadipocytes. Also, selenium promotes embryonic fibroblast cells, and it has a role in adipogenesis (44)

The finding results disagreement with Merrill (45); Barbosa *et al.*(46); Zeng,(26) demonstrates

that selenium induces cell cycle arrest and apoptosis, also disagree with Puspitasari *et al.*(47); Marek *et al.*(48); Valentina *et al.*(49) that have been shown selenium cause decrease cell proliferation *in vitro* and *in vivo*.

These findings also compel the medical research community to continue the search on normal cells such as beta cells, skin cells liver cell and other types of cells especially because of its importance in treating certain diseases.

### Conclusions:

We have carried out the synthesis of new derivatives of nitron and selenonitron, (two selenonitron and three nitrones) in good yield, by the condensation method between carbonyl group and N-monosubstituted hydroxylamines.

The present study revealed that selenonitron ( $N_1$ ) was increased stimulating the cellular proliferation of MAD-MB breast cancer cell line in a concentration and time-dependent manner.

### Acknowledgement:

We greatly acknowledged Prof. Dr. Graham. J. Bodwell, college of science, University of Memorial NL-Canada, for his help to complete this research in his lab and sent the samples for medical test. We are very grateful to Dr. Sherri Christian, Senior Scientist, Beatrice Hunter Cancer Research Institute, Department of Biochemistry Memorial University of Newfoundland- Canada, for doing the medical test.

### Conflicts of Interest: None.

### References:

- Rosselin M, Choteau F, Zeamari K, Nash KM, Das A, Lauricella R, *et al.* Reactivities of substituted alpha-Phenyl -N-tert- butyl Nitrones. *J Org Chem.* 2014; 79: 6615-6626.
- Anderson LL. Diverse applications of nitrones for the synthesis of heterocyclic compounds. *Asian J Org Chem.* 2016;5:9-30.
- Floyd RA, Neto HC, Zimmerman GA, Hensley K, Towner RA. Nitron-based therapeutics for neurodegenerative diseases: Their use alone or in combination with lanthionines. *Free Radic Biol Med.* 2013; 62:145-156.
- Mason RP. Imaging free radicals in organelles, cells, tissue, and *in vivo* with immuno-spin trapping. *Redox Biol.* 2016; 8:422-429.
- Steinbrenner H, Speckmann B, Klotz LO. Selenoproteins: Antioxidant selenoenzymes and beyond. *Arch Biochem Biophys.* 2016;595:113-119.
- Banerjee B, Koketsu M. Recent developments in the synthesis of biologically relevant selenium-containing scaffolds. *Coord Chem Rev.* 2017; 339:104-127.
- Misra S, Boylan M, Selvam A, Spallholz JE, Björnstedt M. Redox-active selenium compounds- From toxicity and cell death to cancer treatment. *Nutrients.* 2016;7: 3536.
- Kieliszek M, Lipinski B, Bła'zejak S. Application of sodium selenite in the prevention and treatment of cancers. *Cells* 2017; 6: 39.
- Lipinski B. Sodium selenite as an anticancer agent. *Anticancer Agents Med Chem.* 2017;17: 658-661.
- Evans SO, Khairuddin PF, Jameson MB. Optimising selenium for modulation of cancer treatments. *Anticancer Res.* 2017; 37 : 6497-6509.
- Li Z, Zhao JB, Sun BZ, Zhou TT, Liu MZ, Liu S, *et al.* Asymmetric nitron synthesis via ligand-enabled copper -catalyzed cope-type hydroamination of cyclopropene with oxime. *J Am Chem Soc.* 2017;139: 11702-11705.
- D'Adamio G, Parmeggiani C, Goti A, Cardona F. Gold supported on silica catalyzes the aerobic oxidation of N, N-disubstituted hydroxylamines to Nitrones. *European J Org Chem.* 2015: 6541-6546.
- Katahara S, Kobayashi S, Fujita K, Matsumoto T, Sato T, Chida N. An iridium-catalyzed reductive approach to nitrones from N-hydroxyamides. *J Am Chem Soc.* 2016; 138: 5246-5249.
- Hao ZQ, Xu B, Gao W, Han YX., Zeng G, Zhang JS. *et al.* Chromium complexes with N,N,N-tridentate quinolinyl anilido-Imine ligand: Synthesis, characterization, and catalysis in ethylene polymerization. *Organometallics* 2015; 34: 2783-2790.
- Pang Y, An B, Lou L, Zhang J, Yan J, Huang L. *et al.* Design, synthesis, and biological evaluation of novel selenium-containing isocombretastatins and phenstatins as antitumor agents. *J Med Chem.* 2017; 60: 7300 -7314.
- Domracheva I, Kanepe-Lapsa I, Jackevica L, Vasiljeva J, Arsenyan P. Selenopheno quinolinones and coumarins promote cancer cell apoptosis by ROS depletion and caspase-7 activation. *factor 2.Life Sci.* 2017; 186: 92-101.
- Domínguez-Álvarez E, Gajdács M, Spengler G, Palop JA, Mar'c MA, Kie'c-Kononowicz K, *et al.* Identification of selenocompounds with promising properties to reverse cancer multidrug resistance. *Bioorg Med Chem Lett.* 2016; 26 : 2821-2824.
- Li W, Guo M, Liu Y, Mu W, Deng G, Li, C. Qiu C. Selenium induces an anti-tumor effect via inhibiting intratumoral angiogenesis in a mouse model of transplanted canine mammary tumor cells. *Biol Trace Elem Res.* 2016;171: 371-379.
- Gajdács M, Spengler G, Sanmartín C, Mar'c MA, Handzlik J, Domínguez-Álvarez E. Selenoesters and selenoanhydrides as novel multidrug resistance reversing agents: A confirmation study in a colon cancer MDR cell line. *Bioorg Med Chem Lett.* 2017; 27 : 797-802.
- Sakalli E, Naziro'glu M, Çi'g B, Övey' I S, Aslan Ko, Sar P. Selenium potentiates the anticancer effect of cisplatin against oxidative stress and calcium ion signaling-induced intracellular toxicity in MCF-7 breast cancer cells: Involvement of the TRPV1



- channel. *J Recept Signal Transduct Res.*2017; 37: 84–93.
21. Falah SF, Saja NM .Essential Trace Elements and Their Vital Roles in Human Body .*Indian J Advances Chem Sci.* 2017; 5(3):127-136
  22. Bozena H , Marta K , Sylvie S, Carlos F, Branislav R ,Thembinkosi D, *et al.* A Summary of new findings on the biological effects of selenium in selected animal species - a critical review. *Int J Mol Sci.* 2017; 18: 2209.
  23. Brozmanova J, Manikova D,Vlckova V, Chovanec M. Selenium: A double-edged sword for defense and offence in cancer. *Arch Toxicol.* 2010; 84:919–938.
  24. Institute of Medicine (IOM) Dietary reference intakes for vitamin C, vitamin E, selenium, and carotenoids. Washington DC: National Academy Press; 2000. 284–324.
  25. John R, Rosa M, Mario J, Stephen B, Richard J. Toxicologic profile for selenium. Atlanta, Georgia GA: USA; 2003.
  26. Zeng H. Selenium as an essential micronutrient: Roles in cell cycle and apoptosis. *Molecules.*2009;14:1263- 1278.
  27. Rayman MP. Selenium and human health. *Lancet.*2012; 11:61452-9.
  28. Vinceti M, Dennert G, Crespi CM, Zwahlen M, Brinkman M, Zeegers MP, *et al.* Selenium for preventing cancer. *Cochrane Database Syst Rev.* 2014 Mar 30; 3 :195
  29. McAuslan BR, Reilly W. Selenium-induced cell migration and proliferation: Relevance to angiogenesis and microangiopathy. *Microvasc Res.* 1986; 32 (1):112-120.
  30. Shin-Hyung P, Jeong-hwan K, Soo-Wan N .Byung-Woo K, GI-Young K, Wun-Jae K, *et al.* .Selenium improves stem cell potency by stimulating the proliferation and active migration of 3T3L1 preadipocytes. *Int J Oncol.* 2014; 44: 336-342
  31. Batool SH. Synthesis of some new selenonitron compounds. *J. Oriental.* 2017; 33 (6): 2821-2826.
  32. Batool SH. Synthesis of new Seleno-Schiff Base Compounds. *J. Der Pharma Chemica.* 2017; 9 (20):25-28.
  33. Veladi P, Girish P, Nageswara R, Vommina V. Synthesis of symmetrical dibenzyl diselenides and disulfides. *J Synthesis.* 2016; 48 (11): 1711-1718.
  34. Maiuolo L, Merino P, Algieri V, Nardi M, Di Gioia ML, Russo B, *et al.* .Nitrones and nucleobase-containing spiro-isoxazolidines derived from isatin and indanone: solvent-free microwave-assisted stereoselective synthesis and theoretical calculations. *Rsc Adv.* 2017; 7: 48980–48988.
  35. Padungros P. Practical synthesis of aromatic dithiocarbamates. *Synthetic Com.* 2014; 44: 2336-2343.
  36. Liu T, Liu Z, Hu D, Wang Y. Synthesis of (Z)-N-arylnitrones . *Synthesis.*2018; 50: 1728-1736.
  37. Cailleau R, Olive M, Cruciger QV. Long-term human breast carcinoma cell lines of metastatic origin preliminary characterization. *In Vitro.* 1978;14(11):911–915.
  38. Srisawat T, Chumkaew P, Heed-Chim W, Sukpondma Y, Kanokwiroon K. Phytochemical screening and cytotoxicity of crude extracts of *Vatica diospyroides* Symington type LS. *Trop J Pharm Res.* 2013; 12:71-6.
  39. Scott M, Lippman MD, Eric A, Klein MD, Phyllis J, Goodman MS. Effect of selenium and vitamin E on risk of prostate cancer and other cancers .The selenium and vitamin E cancer prevention trial (select) *JAMA.* 2009; 301(1):39-51.
  40. Grana X, Reddy EP. Cell cycle control in mammalian cells: role of cyclins, cyclin-dependent kinases (CDKs), growth suppressor genes and cyclin-dependent kinase inhibitors (CKIs). *Oncogenes.*1995; 20: 211- 219.
  41. Kim T, Jung U, Cho DY, Chung AS. S-methylselenocysteine induces apoptosis through caspase activation in HL-60 cells. *Carcinogenesis.* 2001; 22:559-565.
  42. Zeng H. Selenite and selenomethionine promote HL-60 cell cycle progression. *J Nutr.* 2002;132:674-679.
  43. Petra A, Bradley A, Min-Hyuk S, Naranjo S, Xue-Ming X, Yiwen HE, *et al.* The 15kDa selenoprotein and thioredoxin reductase 1 promote colon cancer by different pathways. *J pone.*2015; 10(4): 1-18.
  44. Aishlin E. Selenium treatment promotes adipogenesis in chicken embryonic fibroblasts *In Vitro*, MSC Thesis. USA: Ohio State University. 2013.
  45. Merrill JC. Selenium and Prostate Cancer Prevention: What Next—If Anything? *Cancer Prev Res.*2014; 7(8): 781–785
  46. Barbosa FA, Siminski T, Canto RF, Almeida GM , Mota NS, Ourique F, *et al.* Novel pyrimidinic selenourea induces DNA damage, cell cycle arrest, and apoptosis in human breast carcinoma. *Eur J Med Chem.* 2018; 15(155):503-515
  47. Puspitasari IM, Abdulah R, Yamazaki C, Kameo S, Nakano T, *et al.* Updates on clinical studies of selenium supplementation in radiotherapy. *Radiat Oncol.* 2014; 9: 125
  48. Marek K., Boguslaw L, Stanislaw B. Application of sodium selenite in the prevention and treatment of cancers. *Cells.*2017; 6(4):39
  49. Valentina G, Prajakta K, Jeremy B, Aristi P. Organic selenium compounds as potential chemotherapeutic agents for improved cancer treatment. *Free Radic Biol Med.* 2018 ;127: 80–97

## تحضير وتشخيص مشتقات سيلينونايترين جديدة وتأثيرها على حيوية خط خلايا سرطان الثدي خارج الجسم الحي

كوثر خلف حسن<sup>2</sup>

بتول صالح مهدي حداد<sup>1</sup>

<sup>1</sup> قسم الكيمياء، كلية العلوم، جامعة البصرة، البصرة، العراق.

<sup>2</sup> فرع التشريح البشري، كلية الطب، جامعة البصرة، البصرة، العراق.

### الخلاصة:

تم تحضير مركبات جديدة من النترون والسيلينوننترون. استخدمت طريقه التكاثف بين N-(2- هيدروكسي ايثايل)هيدروكسيل امين ومركبات الكربونيل المعوضه وهي [ البنزل و 4,4- ثنائي كلوروبنزل و 2,2- ثنائي نايتروبنزل] وادت هذه الطريقه الى تحضير مجموعه جديده من مركبات النترون. بينما التكاثف بين N – بنزايل هيدروكسيل امين و مركبات السيلينوكربونيل المعوضه وهي [ثنائي (4- فلوروبنزايل) ثنائي سيلينايد و (4- كلوروبنزايل سيلينونايتريل)] ادت هذه الطريقه الى تحضير مركبات جديده من سيلينوننترون. اما تكاثف N-(4- كلوروفينايل هيدروكسيل امين مع ثنائي بنزايل ثنائي سيلينايد ادى الى تحضير نوع اخر من مركبات سيلينوننترون. شخصت المركبات المحضرة بأستخدام تقنية تحليل العناصر الدقيق والطرائق الطيفية المختلفة كمطيافية تحت الحمراء، مطيافية فوق البنفسجية، مطيافية الرنين النووي المغناطيسي <sup>13</sup>C، <sup>1</sup>H ومطيافية الكتلة. وكدت نتائج التحليل الطيفي وتحليل العناصر الدقيق صحة التراكيب الكيميائية المتوقعة للمركبات المحضرة في هذه الدراسة. كشفت نتائج تأثير مشتقات السيلينونايترين الجديدة (N1) على خلايا سرطان الثدي -MDA MB خارج الجسم الحي زيادة في تكاثر الخلايا بازدياد تراكيز السيلينونايترين وفي كل الفترات الزمنية.

**الكلمات المفتاحية:** حيوية الخلايا، خط خلايا سرطان الثدي MDA- MB-231، نيترونات، الكاثر، سيلينو كربونيل، سيلينونيترونات.