Hemato-Serological Findings as Early signals in Nile Tilapia Oreochromis niloticus Treated with Benzalkonium Chloride

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Abstract

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To evaluate the toxicity of benzalkonium chloride in aquaculture, the hemato-serological indices of Nile tilapia Oreochromis niloticus are used as biomarkers. Following exposure to three concentrations of benzalkonium chloride BAC 0.1, 0.25, 0.50, and 1 mg/l (BAC1,2,3 and 4) in aquaria for two durations 21 and 42 days, the microbiological assay in fish aquaria, in addition to blood parameters were assessed. Except for the mean difference between BAC2 and BAC3 (P > 0.05) at 42 days, the mean values of the bacterial counts revealed a significant difference between all compared groups $(0.05 \ge P \le 0.01)$. Following exposure to the lower concentrations of BAC (1, 2 and 3), the main blood parameters of Oreochromis niloticus namely red blood cells RBCs, and hemoglobin Hb decreased in fluctuated pattern when compared to control treatment (0.05 > P < 0.01), but interestingly peaked at the higher concentration (BAC4), despite there were no significant differences when compared to control (except hemoglobin at 42 days, P< 0.01). Contrarily, the white blood cells WBCs rapidly rose at the first concentration BAC1 (P< 0.01), particularly at the second duration, compared to the control treatments. The counted WBCs changed after exposure to the following two concentrations BAC2, BAC3, before peaking at the higher experimental concentration BAC4 (P < 0.01). The mean cells volume MCV, mean cells haemoglobin MCH and MCHC mean cells haemoglobin concentration exhibited a narrow fluctuation between control and BAC treated fish. Regarding biochemical findings, the average levels of cholesterol and triglycerides showed erratic behavior depending on BAC levels in treatment. The mean alanine transaminase ALT and aspartate aminotransferase AST levels fluctuated between the control and BAC1,2,3 before abruptly increasing at the higher BAC4, with significant differences (0.05 > P < 0.01) when control, BAC1,2,3, compared to BAC4, except AST between BAC3 and BAC4 (insignificant- 42 days). Following BAC exposure, other parameters, including protein and urea remained the same as in control fish. The alteration in some hematoserological markers demonstrated how a compensatory mechanism is generated in response to the stress caused by exposure to biocides.

Keywords: Benzalkonium chloride, Biomarkers, Disinfectant, Oreochromis niloticus, Toxicity.

Introduction

Benzalkonium chloride BAC is a broad-spectrum biocide¹, with the ability to disinfect against fungi, algae, bacteria, and viruses, BAC in lower

concentrations 0.002-0.02% is a preservative included in consumer health products including eye drops and intranasal sprays. On the other hand,

BAC is used for surface disinfection in residential, hospital, and home settings².

The primary mechanism of BAC toxicity is the disruption of the integrity and functionality of the cell membrane, which results in cell death ³. The main sources of BAC released into the environment are sludge and effluent discharged by sewage treatment plants ^{4,5}. In Minnesota, disinfectant quaternary ammonium compounds in wastewater effluent 0.4 μ g/l to 6.6 μ g/l and sediments 0.1 μ g/l to 4.5 µg/l downstream from wastewater treatment discharges have been observed ⁶. These findings are anticipated because the common disposal method for these compounds during their lifetime is down the drain, and although the removal rate in wastewater treatment plants is high, it won't be 100% ⁷. Ecotoxicology studies have shown that BAC is very toxic to several aquatic organisms⁸. Additionally, several quaternary ammonium compounds are combined with other chemical contaminants in the aquatic environment rather than occurring as separate compounds and have been shown to have genotoxic effects in the mammalians, plant, and crustacean cells 9, 10. Experimentally, 16 µg/l BAC resulted in a reduction of the Gramnegative bacterium Acinetobacter baumannii¹¹. Some earlier investigations were undertaken to determine the toxicity of BAC to various fish species, including, for instance, rainbow trout Oncorhynchus mykiss 0.100 to 1.050 mg/l¹², Juvenile Oreochromis niloticus, $LC_{50/96h} = 69.772$ ppm¹³, Cyprinus carpio 1 mg/l for 24, 48, and 96 h ¹⁴. Fish are considered acceptable test organisms because, given their ecological function and trophic location, they serve as significant aqueous pollutant receptors.

Currently, fish serves as a significant and costeffective source of animal protein for almost a billion people worldwide, making up more than

Materials and Methods

The current experiment was carried out at the Aquaculture Lab, Department of Zoology, Faculty of Science, Al-Azhar University (Assiut Branch), Assiut, Egypt. Benzalkonium chloride 50% (LONZAGARD® BKC cGMP), Lonza was used as test chemical for the current assessment. To determine its effects on blood properties, Nile



25% of the total supply ^{15,16}. Water, especially fresh water, is essential for life, but regrettably, it can also contain biological, organic, and inorganic materials which may harm both human and aquatic life ^{17,18}. Today more than ever, efforts are being made to manage water quality to sustain aquaculture's productivity and quality, which has benefitted the economies of many countries ¹⁹. Furthermore, over the past few decades, aquaculture has significantly boosted the output of animal proteins in practically every nation on earth ²⁰. Thereby, fishing pressure and wild stock harvesting from rivers, lakes, oceans, and other open water resources have declined ¹⁵. On the other hand, water quality, environmental stress, the introduction of infectious pathogens, and an increase in the prevalence of fish diseases have all posed several health hazards to aquatic species, particularly fish ²¹. On the one extreme, this has increased the use of various chemicals and aqua medicines in aquaculture to enhance and control fish health and water quality ¹⁵ and minimize production loss on the other extreme ²². Synthetic drugs and chemicals have been used to treat or prevent fish infections with varying degrees of success. However, the risk of their excessive use includes immunosuppression, nephrotoxicity. growth inhibition, the development of drug resistance, environmental issues, and meat residues ²³. To prevent parasite and fungus infections, for instance, copper sulphate, formaldehyde, and malachite green were used in aquaculture as exceptionally powerful traditional disinfectants; however, their usage was linked to several biological and physiological problems ^{24,25}. Tilapias fish is the second-most important cultivated fish worldwide after cyprinids ²⁶. Consequently, the main goal of the current work is to evaluate the safety of BAC in aquaculture practices in terms of the potential health implications of using the Nile tilapia Oreochromis niloticus as an experimental fish.

tilapia (*Oreochromis niloticus*) was exposed to benzalkonium chloride in the current work for 42 days.

Bacteriological assay

After 21, and 42 days, three water samples were collected from each treatment aquarium containing BAC, as well as the control aquarium. The samples

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were collected using clean, sterile, non-reactive caped glass tubes. Samples were sent immediately to the water and wastewater company (Sohag governorate, Egypt) for bacterial enumeration. The bacteria in the water samples were counted using the Pour Plate Method. Heterotrophic plate count (HPC) was used, and the following equation was adopted ²⁷:

Colonies counted

Colony forming units CFU =

The actual volume of sample

Experimental setup and preparation of work solutions (BAC)

Following a week of acclimatization, tilapias with average body weights of 50 ± 7 g and average lengths of 16±4 cm were selected. 150 fish were divided into 5 groups, 3 aquaria $\times 10$ fish per group and transferred to 15 aquaria, 100×40×50 cm. The experiment was planned to run for six weeks, from 5th August to 16th September 2021. Fish were fed commercial pellets during the tests, and water was replaced every 48 hours to remove fecal and nonutilized feed with water that had the same amount of benzalkonium chloride as the siphoned volume. For BAC, a stock solution of 100 mg/l was diluted from the original solution as follows: An amount of $200\mu l = 0.2 \text{ ml} / l \text{ was taken in a volumetric flask}$ 1000 ml, then make up to 1 L with distilled water. The work solution based on the pamphlet on BAC chloride toxicity for freshwater fishes 0.223 - 2.4 mg/l, 96h LC₅₀ was followed for prolonged (chronic) exposure. The work solutions selected are 0.1, 0.25, 0.50 and 1 mg/which are equivalent 100, 250, 500 and 1000 μ l/l, respectively from the stock solution 100 mg/l (preparation based on CAS 8001-54-5; benzalkonium chloride 50% solution, toxicity for freshwater fishes (0.223 - 2.4 mg/L, 96h LC₅₀). In the current study, the control group was singed as C, and the four concentrations (0.1, 0.25, 0.50 and 1 mg/) of BAC were singed as BAC1, BAC2, BAC3, and BAC4, respectively.

Results and Discussion

Bacteriology assay in aquaria

A remarkable decrease (P \leq 0.05), in bacteriology resulted at both experimental durations, Tables 1, 2. In control aquaria, the counted colony forming units were 5023.3±135.4 CFU and 5624.3±270.1 CFU after 21 and 42 days, Baghdad Science Journal

Hematological and Biochemical analysis

Blood samples from the experimental setup were taken at equal intervals for hematological and serological assays, after days 21 and 42 days. In the current investigation, three replicates of each variable were collected. Blood was taken from caudal vein ²⁸ and divided into two parts as follows: To determine the full blood count, the first part was transferred to an EDTA anticoagulant tube. The second portion of the blood sample was put into an Eppendorf tube without EDTA, centrifuged for 10 minutes at 3000 rpm, and the supernatant was then analyzed biochemically. Using an automated technical analyzer (Celltac MEK- 6400J/K), the hemoglobin concentration (Hb), hematocrit (Hct), and red blood cells (RBCs) count of whole blood were measured. White blood cells (WBCs) count, mean corpuscular volume (MCV). mean corpuscular hemoglobin (MCH), mean corpuscular hemoglobin concentration (MCHC) and were determined based on ²⁹. The levels of Aspartate Aminotransferase (AST), Alanine Aminotransferase (ALT), Creatinine, Aspartate Aminotransferase (AST), and Triglyceride were determined in serum using the techniques of ³⁰⁻³².

Statistical analysis

To express the data, the mean and standard variables were tabulated. To deviation of statistically evaluate the impacts of the tested chemical on blood variables between the experimental fish groups, a one-way analysis of variance (ANOVA) test was applied. For data analysis, the SPSS, V: 25 software was used. To compare mean differences, LSD multiple range test was employed. Probability values between 0.05 and 0.01 were evaluated as significant. Statistically nonsignificant, significant, and highly significant outputs were accompanied by symbols NS, * and ** respectively.

respectively. Following that, their numbers rapidly reduced as BAC levels rose. Except for the mean difference between BAC2 and BAC3 (insignificant, P = 0.47) at 42 days, Table 2 has shown the presence of highly significant differences between the recorded mean values, between any compared concentrations including the control group $(0.05 \ge P)$

0.01).

Table 1. Means of bacteriological burdens recorded at the two experimental durations from control and BAC- containing aquaria of Oreochromis niloticus

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and DAC- containing aquaita of Oreochromis hiloticus.										
CFU	С	BAC1	BAC2	BAC3	BAC4					
21days	5023.3 ± 135.4	3511.3±18	3031.6±44.2	2838±61.5	1957.6±24.5					
		7.6								
42 days	5624.3±270.1	3195.6±61.	2703.6±38.7	2652.6±92.	1806.3 ± 65.0					
		9		6						

CFU= colony forming units; C= control; BAC1, BAC2, BAC3, BAC4=0.1,0.25,0.5,1 mg/l benzalkonium chloride, respectively.

Table 2. LSD multiple comparisons for bacteriological colonies between different treatments.

(I - J) Trea	tment (mg/l)	Mean Difference (1-J)	P - value
	After	21 days exposure	
С	BAC1	1512.00 **	.00
	BAC2	1991.66**	.00
	BAC3	2185.33**	.00
	BAC4	3047.66**	.00
BAC1	BAC2	479.66*	.05
	BAC3	673.33**	.00
	BAC4	1535.66**	.00
BAC2	BAC3	193.66**	.01
	BAC4	1056.00**	.00
BAC3	BAC4	862.33**	.00
	After	42 days exposure	
С	BAC1	2428.66**	.01
	BAC2	2920.66**	.01
	BAC3	2971.66**	.01
	BAC4	3818.00**	.00
BAC1	BAC2	492.00**	.00
	BAC3	543.00**	.00
	BAC4	1389.33**	.00
BAC2	BAC3	51.00 ^{NS}	.47
	BAC4	897.33**	.00
BAC3	BAC4	846.33**	.00
CFU= colony j	forming units; C	= control;(BAC1, BAC2, BAC	C3, BAC4) =
(0.1.0.25.0.5.1	mg/l benzalkonii	um chloride, respectively).	

LSD= least significant differences.

The mean difference is significant at level 0.05.

The mean difference is significant at level 0.01.

^{NS} The mean difference is nonsignificant.

Hematological parameters

The data in Table 3 summarizes the average blood indices measured during the experiment involving Nile tilapia Oreochromis niloticus exposure to benzalkonium chloride. When compared to the control group, the mean RBCs and hemoglobin exhibited decreased variation (0.05> P <0.01) following the exposure to BAC1,2 and 3, then peaked at the higher concentration BAC4 at both sampling times 21 and 42 days compared to control groups, despite the presence of no statistical differences (P > 0.05) as indicated in Table 4. In relation to BAC concentration, MCV varied; it peaked at BAC4, compared to control fish for both 21 and 42 days, respectively (P < 0.01) as shown in table 4. However, there was an erratic fluctuation between the two treatments, namely control and BAC4. At the lower BAC1,2,3 concentrations, MCH increased considerably with comparable differences, particularly at 21 days ($0.05 > P \le 0.01$), while at the higher BAC4 concentration it decreases once again at both time periods, Table 3. MCHC exhibited a dramatic decrease at both periods between the BAC1,2,3 compared to control means (P <0.01). WBCs varied between the control and the three concentrations BAC1,2,3, before rising



sharply at BAC4 (P <0.01) at both sampling intervals 21 and 42 days. All scored hematological parameters were concentration-dependent rather than time-dependent. Whoever, WBCs were shown

to be influenced by both BAC concentration and exposure time. The mean values MCV and MCH were noticed to be out of balance during the BAC exposure period, Tables 3, 4.

 Table 3. Means of hematological parameters recorded at the two experimental durations from control and BAC- containing aquaria of *Oreochromis niloticus*.

Parameters (un	it)	C	BAC1	BAC2	BAC3	BAC4		
RBCs (×10 ⁶ /µl)	21 days	2.2±0.3	1.7 ± 0.1	1.8 ± 0.07	1.8 ± 0.1	2.5 ± 0.07		
	42 days	2.3±0.1	1.8 ± 0.09	1.6 ± 0.1	1.5±0.3	2.3 ± 0.07		
Hb (g/dl)	21 days	7.8±0.2	6.7±0.7	6.7±0.3	7.5 ± 0.8	9.8±0.2		
	42 days	8.8±0.2	8.0±0.3	7.1±1.3	6.9±1.03	10±0.2		
MCV (fL)	21 days	120±1.7	156.6 ± 20.5	173.6±6.7	155±4.4	166.6 ± 5.4		
	42 days	117.6±1.8	142 ± 10.1	156.3±7.5	187.6 ± 4.5	163.3±20		
MCH (pg)	21 days	34.6±1.9	37.9 ± 1.8	37.9±1.3	41±1.3	24.06±0.9		
	42 days	37.0±2.4	42.3±4.6	40.8 ± 2.5	44.3±2.05	42.2±1.7		
MCHC (g/dL)	21 days	33.3±2.4	24.5±3.0	21.7±0.7	26.5 ± 0.08	24.4±0.9		
	42 days	36.4±0.9	27.5 ± 3.04	23.1±1.5	23.6±1.6	26.1±2.7		
Hct (%)	21 days	34.2 ± 2.01	27.8 ± 5.5	30.8±2.3	28.1±2.9	40.9±1.1		
	42 days	35.5±2.1	28.8 ± 5.4	32.0 ± 7.8	30.1±6.6	38.9±4.9		
WBCs (×10 ³ /µl)	21 days	27.6±1.1	50.5±13.9	45.2±11.6	59.3±7.7	107±7.9		
	42 days	36.6±4.7	71.0±1.6	94.7±9.5	48.4±9.5	106.8±12.9		
C= control;(BAC1, BAC2, BAC3, BAC4) = (0.1,0.25,0.5,1 mg/l benzalkonium chloride, respectively).								

Table 4. LSD multiple comparisons for hematological parameters between different treatments and

	_	_			time	s. –					
Parameter	rs	С	С	С	С	BAC	BAC	BAC	BAC	BAC	BAC
		&	&	&	&	1	1	1	2	2	3
						&	&	&	&	&	&
		BAC									
		1	2	3	4	2	3	4	3	4	4
RBCs	21 d	$.02^{*}$.03*	$.04^{*}$.08 ^{NS}	.80 ^{NS}	.65 ^{NS}	$.00^{**}$.83 ^{NS}	$.00^{**}$	$.00^{**}$
	42 d	.03*	$.00^{**}$	$.00^{**}$.63 ^{NS}	.28 ^{NS}	.13 ^{NS}	$.01^{**}$.61 ^{NS}	$.00^{**}$	$.00^{**}$
Hb	21 d	.03*	.03*	.55 ^{NS}	$.00^{**}$	1.0^{NS}	.10 ^{NS}	$.00^{**}$.10 ^{NS}	$.00^{**}$	$.00^{**}$
	42 d	.26 ^{NS}	.03*	$.02^{*}$.12 ^{NS}	.25 ^{NS}	.14 ^{NS}	$.01^{**}$.69 ^{NS}	$.00^{**}$	$.00^{**}$
MCV	21 d	$.00^{**}$	$.00^{**}$	$.00^{**}$	$.00^{**}$.09 ^{NS}	.86 ^{NS}	.30 ^{NS}	.07 ^{NS}	.47 ^{NS}	.23 ^{NS}
	42 d	.03*	$.00^{**}$	$.00^{**}$	$.00^{**}$.18 ^{NS}	$.00^{**}$	$.05^{*}$	$.01^{**}$.50 ^{NS}	.03*
MCH	21 d	.03*	.08 ^{NS}	$.00^{**}$	$.00^{**}$.55 ^{NS}	$.04^{*}$	$.01^{**}$	$.01^{**}$	$.00^{**}$.43 ^{NS}
	42 d	.08 ^{NS}	.19 ^{NS}	$.02^{*}$.08 ^{NS}	.59 ^{NS}	.48 ^{NS}	.96 ^{NS}	.23 ^{NS}	.62 ^{NS}	.45 ^{NS}
MCHC	21 d	$.00^{**}$	$.00^{**}$	$.00^{**}$	$.00^{**}$.12 ^{NS}	.26 ^{NS}	.58 ^{NS}	$.017^{**}$	$.04^{*}$.54 ^{NS}
	42 d	$.00^{**}$	$.00^{**}$	$.00^{**}$	$.00^{**}$	$.04^{*}$.07 ^{NS}	.49 ^{NS}	.79 ^{NS}	.14 ^{NS}	.22 ^{NS}
Hct	21 d	$.05^{*}$.26 ^{NS}	$.06^{NS}$	$.04^{*}$.32 ^{NS}	.91 ^{NS}	$.00^{**}$.37 ^{NS}	$.00^{**}$	$.00^{**}$
	42 d	.23 ^{NS}	.52 ^{NS}	.32 ^{NS}	.52 ^{NS}	.55 ^{NS}	.81 ^{NS}	.08 ^{NS}	.71 ^{NS}	.21 ^{NS}	.12 ^{NS}
WBCs	21 d	.09 ^{NS}	.19 ^{NS}	.03*	$.00^{**}$.68 ^{NS}	.49 ^{NS}	$.00^{**}$.28 ^{NS}	$.00^{**}$	$.00^{**}$
	42 d	$.00^{**}$	$.00^{**}$.16 ^{NS}	$.00^{**}$.01**	.01**	$.00^{**}$	$.00^{**}$.15 ^{NS}	$.00^{**}$

C= control;(BAC1, BAC2, BAC3, BAC4) = (0.1,0.25,0.5,1 mg/l benzalkonium chloride, respectively). * The mean difference is significant at level 0.05.

** The mean difference is significant at level 0.01.

^{NS} The mean difference is nonsignificant.

Serological analysis

In *Oreochromis niloticus*, the difference in the mean values of total protein and urea between the control groups and the BAC-exposed group was not statistically significant (P>0.05), Tables 5,6. The mean glucose level rose in the blood of fish exposed

to BAC1,2,3 compared to the control fish group (P<0.01), particularly at the 21 days, then increased proportionally before declining once more at BAC4, at days 21 with no significant differences. However, the mean glucose differed significantly at 42 days between BAC2 & BAC4 (P<0.01), and BAC3,



BAC4 (P=0.01). The levels of cholesterol were measured throughout the two test durations in a zigzag pattern; in comparison to the control group, the levels of cholesterol determined from BAC1 were higher at both test durations 21 and 42 days, when compared to control findings (P<0.01), Tables 5,6. Following that, cholesterol started to decline again at the second exposure BAC2. Finally, cholesterol rose at the third treatment BAC3, and reduced at the higher exposure concentration BAC4. As well as cholesterol, triglyceride mean levels were upper-lower in trend and were time -BAC concentrations dependent. The means ALT and AST levels showed a fluctuating pattern between the control and the concentrations of BAC1,2,3, before drastically peaking at the higher concentration BAC4 at 21 and 42 days, showing different degrees of significance $(0.05>P\leq0.01)$. When compared to the starting level of the control group, the creatinine level was found to have increased significantly after the experiment $(0.05>P\leq0.01)$.

 Table 5. Means of serological parameters recorded at the two experimental durations from control and BAC- containing aquaria of *Oreochromis niloticus*.

d	inu DAC- Co	ntanning ayt	ial la Ul Uleu		cus.			
Parameters (unit)		С	BAC1	BAC2	BAC3	BAC4		
	21 days	3.1±0.04	3.2±0.1	3.1±0.02	3.1±0.1	3.1±0.01		
Protein (mg/dl)	42 days	3.02±0.04	3.01±0.1	2.9±0.1	2.9±0.05	3.01±0.09		
	21 days	92±3.5	115.3±3.1	134±12.4	135.3±4.5	123.6±5.8		
Glucose (mg/dl)	42 days	105.3 ± 4.5	118.3±21.1	111.3 ± 5.08	119.6±4.03	149 ± 7.09		
	21 days	170.3 ± 4.0	192.3±6.7	185±12.0	200±3.5	182.3 ± 7.1		
Cholesterol (mg/dl)	42 days	152.3±2.7	195±10.7	162.6±11.8	181.3±7.2	BAC3BAC4 3.1 ± 0.1 3.1 ± 0.01 2.9 ± 0.05 3.01 ± 0.09 135.3 ± 4.5 123.6 ± 5.8 119.6 ± 4.03 149 ± 7.09 200 ± 3.5 182.3 ± 7.1 181.3 ± 7.2 192.3 ± 6.7 128.3 ± 7.6 165.3 ± 4.5 125 ± 4.4 155.3 ± 9.8 0.4 ± 0.01 0.5 ± 0.02 0.6 ± 0.01 0.7 ± 0.05 15 ± 2.3 14 ± 0.8 14.6 ± 1.3 14.6 ± 2.2 25.3 ± 4.03 33.6 ± 3.7 38 ± 1.7 62.6 ± 11.1 45.6 ± 4.9 154 ± 6.9 178.6 ± 10.7 189 ± 10.3		
	21 days	98.6±3.6	64.6±4.9	131.6±46.5	128.3±7.6	165.3 ± 4.5		
l rigiyceride (mg/di)	42 days	121±2.3	$\begin{array}{rrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrrr$	125±4.4	155.3±9.8			
	21 days	0.5 ± 0.01	0.5 ± 0.02	0.4 ± 0.03	0.4 ± 0.01	0.5 ± 0.02		
Creatinine (mg/di)	42 days	0.5 ± 0.02	0.5 ± 0.02	C1BAC2BAC3BAC4 ± 0.1 3.1 ± 0.02 3.1 ± 0.1 3.1 ± 0.01 ± 0.1 2.9 ± 0.1 2.9 ± 0.05 3.01 ± 0.09 ± 3.1 134 ± 12.4 135.3 ± 4.5 123.6 ± 5.8 ± 21.1 111.3 ± 5.08 119.6 ± 4.03 149 ± 7.09 ± 6.7 185 ± 12.0 200 ± 3.5 182.3 ± 7.1 ± 10.7 162.6 ± 11.8 181.3 ± 7.2 192.3 ± 6.7 ± 4.9 131.6 ± 46.5 128.3 ± 7.6 165.3 ± 4.5 5 ± 18 153.3 ± 13.6 125 ± 4.4 155.3 ± 9.8 0.02 0.4 ± 0.03 0.4 ± 0.01 0.5 ± 0.02 0.02 0.6 ± 0.01 0.6 ± 0.01 0.7 ± 0.05 ± 1.8 13 ± 1.7 15 ± 2.3 14 ± 0.8 $:0.8$ 14 ± 0.8 14.6 ± 1.3 14.6 ± 2.2 ± 2.7 25 ± 3.8 25.3 ± 4.03 33.6 ± 3.7 ± 6.7 39.6 ± 2.2 38 ± 1.7 62.6 ± 11.1 ± 4.03 49.6 ± 7.6 45.6 ± 4.9 154 ± 6.9 ± 11.3 143.3 ± 6.2 178.6 ± 10.7 189 ± 10.3 ng/l $henzalkonium chloride, respectively).henzalkonium chloride, respectively$				
	21 days	14.6±1.3	14.6±1.8	13±1.7	15 ± 2.3	14 ± 0.8		
Urea (mg/dl)	42 days	15±0.8	13±0.8	14 ± 0.8	14.6±1.3	14.6 ± 2.2		
	21 days	26.3±2.8	25.6±2.7	25±3.8	25.3 ± 4.03	33.6±3.7		
ALI(U/L)	42 days	39.6±2.2	37.3±6.7	39.6±2.2	38±1.7	62.6±11.1		
	21 days	38.3±1.8	42.6±4.03	49.6±7.6	45.6±4.9	154±6.9		
ASI(U/L)	42 days	125.3±6.5	105±11.3	143.3±6.2	178.6±10.7	189±10.3		
$C = control \cdot (BAC1, BAC2, BAC3, BAC4) = (0, 1, 0, 25, 0, 5, 1, mg/l, benzalkonium chloride, respectively)$								

 Table 6. LSD multiple comparisons for serological parameters between different treatments and time.

Parameters		C	С	С	C	BAC	BAC	BAC	BAC	BAC	BAC
		&	&	&	&	1	1	1	2	2	3
						&	&	&	&	&	&
		BAC	BAC	BAC							
		1	2	3	4	2	3	4	3	4	4
Protein	21 d	.20 ^{NS}	.80 ^{NS}	.74 ^{NS}	.90 ^{NS}	.29 ^{NS}	.12 ^{NS}	.24 ^{NS}	.56 ^{NS}	.90 ^{NS}	.65 ^{NS}
	42 d	.89 ^{NS}	.78 ^{NS}	.33 ^{NS}	.94 ^{NS}	.89 ^{NS}	.40 ^{NS}	.94 ^{NS}	.47 ^{NS}	.83 ^{NS}	.36 ^{NS}
Glucose	21 d	$.00^{**}$	$.00^{**}$	$.00^{**}$	$.00^{**}$	$.01^{**}$	$.00^{**}$.20 ^{NS}	.83 ^{NS}	.12 ^{NS}	.09 ^{NS}
	42 d	.20 ^{NS}	.54 ^{NS}	.16 ^{NS}	$.00^{**}$.48 ^{NS}	.89 ^{NS}	.01**	.40 ^{NS}	$.00^{**}$	$.01^{**}$
Cholesterol	21 d	$.00^{**}$	$.05^{*}$	$.00^{**}$.10 ^{NS}	.30 ^{NS}	.28 ^{NS}	.16 ^{NS}	$.04^{*}$.70 ^{NS}	$.02^{*}$
	42 d	$.00^{**}$.21 ^{NS}	$.00^{**}$	$.00^{**}$	$.00^{**}$.10 ^{NS}	.73 ^{NS}	.03*	$.00^{**}$.18 ^{NS}
Triglyceride	21 d	.11 ^{NS}	.12 ^{NS}	.15 ^{NS}	$.00^{**}$	$.00^{**}$	$.00^{**}$	$.00^{**}$.86 ^{NS}	.11 ^{NS}	.08 ^{NS}
	42 d	.36 ^{NS}	.01**	.70 ^{NS}	$.00^{**}$	$.05^{*}$.59 ^{NS}	.03*	$.02^{*}$.85 ^{NS}	$.01^{**}$
Creatinine	21 d	$.00^{**}$	$.02^{*}$.24 ^{NS}	$.01^{**}$	$.00^{**}$	$.00^{**}$.76 ^{NS}	.19 ^{NS}	$.00^{**}$	$.00^{**}$
	42 d	1.0^{NS}	$.00^{**}$.10 ^{NS}	$.00^{**}$	$.00^{**}$.10 ^{NS}	$.00^{**}$	$.02^{*}$.08 ^{NS}	$.00^{**}$
Urea	21 d	1.0^{NS}	.31 ^{NS}	.83 ^{NS}	.68 ^{NS}	.31 ^{NS}	.83 ^{NS}	.68 ^{NS}	.23 ^{NS}	.54 ^{NS}	.54 ^{NS}
	42 d	.14 ^{NS}	.44 ^{NS}	.79 ^{NS}	.79 ^{NS}	.44 ^{NS}	.21 ^{NS}	.21 ^{NS}	.60 ^{NS}	.60 ^{NS}	1.0^{NS}
ALT	21 d	.83 ^{NS}	.68 ^{NS}	.76 ^{NS}	$.04^{*}$.83 ^{NS}	.91 ^{NS}	.03*	.91 ^{NS}	$.02^{*}$	$.02^{*}$
	42 d	.68 ^{NS}	1.0^{NS}	.76 ^{NS}	$.00^{**}$.68 ^{NS}	.90 ^{NS}	$.00^{**}$.76 ^{NS}	$.00^{**}$	$.00^{**}$
AST	21 d	.40 ^{NS}	$.04^{*}$.17 ^{NS}	$.00^{**}$.19 ^{NS}	.56 ^{NS}	$.00^{**}$.443 ^{NS}	$.00^{**}$	$.00^{**}$
	42 d	.15 ^{NS}	.20 ^{NS}	.00**	.00**	.01**	.00**	.00**	.02*	.00**	.45 ^{NS}



C= control;(BAC1, BAC2, BAC3, BAC4) = (0.1,0.25,0.5,1 mg/l benzalkonium chloride, respectively).

The mean difference is significant at level 0.05.

** The mean difference is significant at level 0.01.

^{NS} The mean difference is nonsignificant.

Discussion

The ability of BAC to alter cell functions and damage cell membranes has been suggested as the mechanism by which it can be toxic³. Petroleum derivatives, insecticides, nanoparticles, cosmetics, pharmaceuticals, and health and personal care products are just a few of the many synthetic and semi-synthetic substances produced every day to meet humanity's insatiable demand for industrialization and development 33,34, but their negative effects are unavoidable. If this strategy is not controlled and managed, it will eventually cause major issues for both people and their environment. For instance, the research by ³⁴ has demonstrated the ability of *Phaseolus vulgaris* L. to absorb three drugs from soil: diclofenac, mefenamic acid, and metronidazole. A significant challenge facing the entire world today is the entry of significant amounts of produced pollutants from point and nonpoint sources into natural ecosystems. The process of disposal for products containing quaternary ammonium compounds after use is often "downthe-drain" to wastewater treatment systems; as a result, the aquatic environment has the highest likelihood of environmental impact⁷. Fish are consumed throughout the world because they are high in protein and long-chain n-3 polyunsaturated fatty acids ³⁵. Consequently, maintaining the quality of fish is a big concern, and over time, several preservatives have been used to address this problem. As а disinfectant. formalin at concentrations between 15 - 25 mg/l was effective to preserve fish in the past times. But, the best use of formalin levels has, however, been violated frequently. Although biocides are widely used in different industries, they are largely many uncontrolled and as a result, pose an environmental risk ³⁶, thus their stewardship is essential and critical ³⁷. Previous works have demonstrated the effects of BAC on both animals and humans. For example, the 96 h LC₅₀ for BAC 69.772 ppm revealed negative impacts on the experimental juvenile Oreochromis *niloticus* ¹³. On the other hand, human respiratory epithelial cells exposed to BAC at amounts normally found in commercially available nasal

solutions were shown to significantly modify their DNA in vitro, raising the possibility that apoptosis and necrosis have occurred ³⁸. The drastic decrease in bacteriology count resulted here confirming the efficacy of BAC as a disinfectant. Bacteria multiply quickly, infect fish, and increase mortality rates in a few days or weeks ³⁹. Using a time-to-reduce assay, the disinfecting efficacy of BAC on microbiological colonies in the experimental aquaria elucidated the severe impact of BAC ($0.05 \ge P \le 0.01$) on the bacterial count. Similar to the current observation, the growth of alga *Pseudokirchneriella subcapitata* was inhibited after exposure to 0.25 mg/l BAC /72 hours ¹⁰. In consistence, study ¹¹ on the bacteria Acinetobacter baumannii, BAC, on the one hand, hindered gentamicin killing and, on the other hand, dramatically increased the rate at which resistance mutants appeared. Regarding haematological cells biomarkers, blood serve as excellent biomarkers of biological, physical, or chemical changes in the fish's environment ^{40, 41}. Alterations in hematological parameters are thus reliable indicators of deterioration in water quality 42,43 caused by xenobiotics. WBCs levels have fluctuated in the current experiment because the immune system was attempting to control stress, particularly as the microbial burden was lowering. Following exposure to the first three BAC1.2.3 concentrations, red blood cells declined, then increased to levels comparable to control fish at BAC4 at both exposure times. This pattern of RBCs fluctuation is in accordance with some similar studies on Labeo rohita exposed to fungicide propiconazole⁴⁴, and Oncorhynchus mykiss exposed to pesticides ⁴⁵. Fish exposed to higher BAC4 exhibited higher hematocrit levels than control fish, which may mean that hematocrit is increased simultaneously with increasing RBCs, which may be produced in high numbers to compensate for loss. Furthermore, serological indices such as total glucose, triglycerides, ALT, protein. AST. creatinine, urea, and cholesterol have all been employed as responsive biomarkers to evaluate the influence of stressors in both field and laboratory investigations ⁴⁶⁻⁴⁸. For example, toxic stress is known to cause fish to experience transient changes in blood glucose that may level out at a new homeostatic level 49,50. The significant rise in glucose levels, especially at the highest BAC4, may imply the process of glycogenolysis to fulfill the energy requirements necessitated by toxic stress or by a reduction in glycogen synthesis as a reaction to BAC-mediated stress. Oreochromis niloticus, exposed to alkyl benzene sulfonate caused neurological control posing fish to leap out (hyperactivities) of a treatment aquaria⁵¹. Similarly, two fish species Cirrhina mrigala and Puntius sophore showed altered carbohydrate metabolism after being exposed to 0.005 mg/l of alkyl benzene sulphonate ⁵². Since the primary function of the liver is to maintain lipid homeostasis, changes in blood levels of cholesterol and triglycerides signify liver dysfunction ⁵³. Triglyceride and cholesterol levels were found to fluctuate erratically in the current investigation, which is likely due to sterol biosynthesis dysfunction. The experiments on Oreochromis niloticus exposed to benzene sulphonate resulted in elevated levels of cholesterol and triglycerides ⁵¹, which were attributed to the disruption in fat metabolism, which was consistent with the current explanations. Similar investigations using various fish subjected to different types of environmental toxicants have all got results that are consistent with the current findings. For instance, Oncorhynchus mykiss, following exposure to phenol

Conclusion

This assessment is the first to evaluate the impact of widely applicable quaternary ammonium compound (benzalkonium chloride) on Nile tilapia *Oreochromis niloticus* in Egypt. The current findings have revealed that BAC has toxic potential, which resulted in a bacterial count. Furthermore, the main hematological and serological parameters responded to BAC in a concentration-dependent manner. Furthermore, the main hematological and serological parameters responded to BAC in a

Author's Declaration

- Conflicts of Interest: None.
- We hereby confirm that all the Figures and Tables in the manuscript are ours. Furthermore, any Figures and images that are not ours have been included with the necessary permission for re-publication, which is attached to the



⁵⁴, Oreochromis niloticus exposed to copper sulphate ⁴⁷, Oreochromis niloticus sampled from a contaminated environment 46,55. As a stressor, BAC is expected to cause physiological changes associated with the production of free radicals, causing oxidative stress on the biosynthesis within the animal body. BAC was suggested to pass through the blood-placental barrier and enter the mouse neonatal brain ⁵⁶, generating physiological alterations. The remarkable patterns of ALT and AST found here suggest that they are experiencing stress brought on by BAC exposure, particularly at BAC4 exposure compared to control findings (0.05>P<0.01). In accordance with the literature, ALT and AST are found in a variety of tissues and cells, and structural injury to the liver causes them to be discharged into the blood in large quantities ⁵⁷. Similar to the current findings, elevated AST and ALT activities were observed in the blood and liver tissues of numerous fish species subjected to varied pesticide doses for various periods of time ⁵⁸. Also, the study on Cyprinus carpio exposed to BAC at a similar level to the maximum concentration in the current study, resulted in a significant drop in antioxidant enzymes, indicating liver impairment ¹⁴. Our interpretations of AST and ALT are similarly consistent with those of ⁵⁹, who suggested that the fluctuating levels represent an attempt by fish to achieve balance while being exposed to biochemical and physiological function – impairing chemicals.

concentration-dependent manner $(0.05 \ge P \le 0.01)$. Consequently, there is a possibility that BAC could reach the environment and aquatic ecosystems through treated and untreated sewage, including domestic, industrial, and urban runoff because of its extensive use in disinfection. However, the current research suggests that the recommended BAC save concentration for freshwater fish is between 0.1 and 0.5ppm.

manuscript. The author has signed an animal welfare statement.

- Ethical Clearance: The project was approved by the local ethical committee in Department of Zoology, Faculty of Science, Al- Azhar University (Assiut Branch), 71524 Assiut, Egypt. The authors confirm that, no environmental or health threats have been

Author's Contribution Statement

M. A. E.: Prepared fish samples and setup experiments; M. M.: Supervision of the research and revised it; A. B..: Prepared chemical solutions,

References

- 1. Pereira BMP, Tagkopoulos I. Benzalkonium Chlorides: Uses, Regulatory Status, and Microbial Resistance. *Appl Environ* Microbiol. 2019; 85(13): e00377-00319. <u>https://doi.org/10.1128/AEM.00377-19</u>
- 2. Russo C, Kundi M, Lavorgna M, Parrella A, Isidori M. Benzalkonium chloride and anticancer drugs in binary mixtures: reproductive toxicity and genotoxicity in the freshwater crustacean Ceriodaphnia dubia. Arch Environ Contam Toxicol. 2018 546-556. 74(4): https://doi.org/10.1007/s00244-017-0473-y
- Christen V, Faltermann S, Brun NR, Kunz PY, Fent K. Cytotoxicity and molecular effects of biocidal disinfectants (quaternary ammonia, glutaraldehyde, poly (hexamethylene biguanide) hydrochloride PHMB) and their mixtures in vitro and in zebrafish eleuthero-embryos. Sci Total Environ. 2017; 586: 1204-1218.
 - https://doi.org/10.1016/j.scitotenv.2017.02.114
- 4. He Z-W, Liu W-Z, Tang C-C, Liang B, Guo Z-C, Wang L, et al. Performance and microbial community responses of anaerobic digestion of waste activated sludge to residual benzalkonium chlorides. Energy Convers Manag. 2019 ; 202: 112211.

https://doi.org/10.1016/j.enconman.2019.112211

- Barber OW, Hartmann EM. Benzalkonium chloride: A systematic review of its environmental entry through wastewater treatment, potential impact, and mitigation strategies. Crit Rev Environ Sci Technol. 2022; 52(15): 2691-2719. https://doi.org/10.1080/10643389.2021.1889284
- Pati SG, Arnold WA. Comprehensive screening of quaternary ammonium surfactants and ionic liquids in wastewater effluents and lake sediments. Environ Sci Process Impacts 2020; 22(2): 430-441. <u>http://doi.org/10.1039/c9em00554d</u>
- DeLeo PC, Huynh C, Pattanayek M, Schmid KC, Pechacek N. Assessment of ecological hazards and environmental fate of disinfectant quaternary ammonium compounds. Ecotoxicol Environ Saf. 2020; 206(2020): 111116. http://doi.org/10.1016/j.ecoenv.2020.111116
- 8. Kim T-K, Jang M, Hwang YS. Adsorption of benzalkonium chlorides onto polyethylene

aquaria and performed the statistics; and R. S.: suggestion the research, writing - review and editing.

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microplastics: Mechanism and toxicity evaluation. J Hazard Mater. 2022; 426: 128076. https://doi.org/10.1016/j.jhazmat.2021.128076

- Hora PI, Pati SG, McNamara PJ, Arnold WA. Increased Use of Quaternary Ammonium Compounds during the SARS-CoV-2 Pandemic and Beyond: Consideration of Environmental Implications. Environ Sci Technol Lett. 2020; 7(9): 622-631. <u>https://doi.org/10.1021/acs.estlett.0c00437</u>
- Elersek T, Zenko M, Filipic M. Ecotoxicity of disinfectant benzalkonium chloride and its mixture with antineoplastic drug 5-fluorouracil towards alga Pseudokirchneriella subcapitata. Peer J. 2018; 6: e4986. http://doi.org/10.7717/peerj.4986
- Short FL, Lee V, Mamun R, Malmberg R, Li L, Espinosa MI, et al. Benzalkonium chloride antagonises aminoglycoside antibiotics and promotes evolution of resistance. EBio Medicine 2021; 73: 103653. <u>http://doi.org/10.1016/j.ebiom.2021.103653</u>
- 12. Antunes SC, Nunes B, Rodrigues S, Nunes R, Fernandes J, Correia AT. Effects of chronic exposure to benzalkonium chloride in Oncorhynchus mykiss: cholinergic neurotoxicity, oxidative stress, peroxidative damage and genotoxicity. Environ Toxicol Pharmacol. 2016; 45: 115-122. https://doi.org/10.1016/j.etap.2016.04.016
- Ikisa K, Babatunde B, Hart A. Acute toxicity of benzalkonium chloride mixture with treated produced water to juveniles of freshwater tilapia-Oreochromis niloticus. J Appl Sci Enviro Manag. 2019; 23(6): 1169-1174. http://doi.org/10.4314/jasem.v23i6.26
- Gheorghe S, Mitroi DN, Stan MS, Staicu CA, Cicirma M, Lucaciu IE, et al. Evaluation of Sub-Lethal Toxicity of Benzethonium Chloride in Cyprinus carpio Liver. Appl Sci. 2020; 10(23): 8485. https://doi.org/10.3390/app10238485
- Mishra S, Das R, Das B, Choudhary P, Rathod R, Giri B, et al. Status of Aqua-medicines, drugs and chemicals use in India: A Survey Re-port. J Aquac Fish. 2017; 1(004). <u>http://doi.org/10.24966/AAF-5523/100004</u>
- 16. Khan S, Rehman A, Shah H, Aadil RM, Ali A, Shehzad Q, et al. Fish Protein and its derivatives: The novel applications, bioactivities, and their



current

study.

2023, 20(3 Suppl.): 945-956 https://dx.doi.org/10.21123/bsj.2023.7961 P-ISSN: 2078-8665 - E-ISSN: 2411-7986 Baghdad Science Journal

functional significance in food products. Food Rev Int. 2022 ; 38(8): 1607-1634. https://doi.org/10.1080/87559129.2020.1828452

- 17. Alsaeed RD, Alaji B, Ibrahim M. Modeling Jar Test Results Using Gene Expression to Determine the Optimal Alum Dose in Drinking Water Treatment Plants. Baghdad Sci J. 2022 ; 19(5): 951-965. <u>https://doi.org/10.21123/bsj.2022.6452</u>
- Said REM, Said AS, Saber SAL, ElSalkh BAE. Biomarker Responses in Sclerophrys regularis (Anura: Bufonidae) Exposed to Atrazine and Nitrate. Pollution 2022; 8(4): 1387-1397. <u>https://doi.org/10.22059/POLL.2022.339894.1386</u>
- Hu Z, Li R, Xia X, Yu C, Fan X, Zhao Y. A method overview in smart aquaculture. Environ Monit Assess. 2020; 192(8): 493. https://doi.org/10.1007/s10661-020-08409-9
- Jahangiri L, Esteban MÁ. Administration of probiotics in the water in finfish aquaculture systems: a review. Fishes 2018; 3(3): 33. <u>https://doi.org/10.3390/fishes3030033</u>
- Nathanailides C, Kolygas M, Choremi K, Mavraganis T, Gouva E, Vidalis K, et al. Probiotics Have the Potential to Significantly Mitigate the Environmental Impact of Freshwater Fish Farms. Fishes 2021; 6(4): 76. <u>https://doi.org/10.3390/fishes6040076</u>
- Assefa A, Abunna F. Maintenance of fish health in aquaculture: review of epidemiological approaches for prevention and control of infectious disease of fish. *Vet Med Int.* 2018; 2018: 10 https://doi.org/10.1155/2018/5432497
- Kang H-S, Lee S-B, Shin D, Jeong J, Hong J-H, Rhee G-S. Occurrence of veterinary drug residues in farmed fishery products in South Korea. Food Control. 2018; 85: 57-65. <u>https://doi.org/10.1016/j.foodcont.2017.09.019</u>
- Leal JF, Neves MGP, Santos EB, Esteves VI. Use of formalin in intensive aquaculture: properties, application and effects on fish and water quality. *Rev Aquac*. 2018; 10(2): 281-295. https://doi.org/10.1111/raq.12160
- Gallani SU, Valladão GMR, Assane IM, Alves LdO, Kotzent S, Hashimoto DT, et al. Motile Aeromonas septicemia in tambaqui Colossoma macropomum: Pathogenicity, lethality and new insights for control and disinfection in aquaculture. Microb Pathog. 2020 ; 149: 104512. https://doi.org/10.1016/j.micpath.2020.104512
- 26. FAO. The State of World Fisheries and Aquaculture 2018 - Meeting the sustainable development goals. Rome. Licence: CC BY-NC-SA 3.0 IGO. 2018 <u>https://www.fao.org/3/i9540en/i9540en.pddf</u>
- 27. Lavorgna M, Russo C, D'Abrosca B, Parrella A, Isidori M. Toxicity and genotoxicity of the

quaternary ammonium compound benzalkonium chloride (BAC) using Daphnia magna and Ceriodaphnia dubia as model systems. Environ Pollut. 2016; 210: 34-39. https://doi.org/10.1016/j.envpol.2015.11.042

 Farrag MMS, Said RE, Elmileegy IMH, Abou Khalil NS, Abdel allah ESA, El-Sawy MF, et al. Effects of penconazole and copper nanoparticle fungicides on redbelly tilapia, Coptodon zillii (Gervais, 1848): Reproductive outcomes. Int J Aquat Biol. 2022; 9(6): 370-382.

http://doi.org/10.22034/ijab.v9i6.1196

- 29. Bhatnagar N. Dacie and Lewis Practical Haematology (12th edition) By B. J. Bain, I. Bates and M. A. Laffan, Elsevier, London, 2017. Br J Haematol. 2017; 178(4): 652. https://doi.org/10.1111/bjh.14872
- Reitman S, Frankel S. A colorimetric method for determination of serum aspartate and alanine aminotransferases. Am J Clin Pathol. 1975; 28(1): 56. <u>https://doi.org/10.1093/ajcp/28.1.56</u>
- Trinder P. Determination of glucose in blood using glucose oxidase with an alternative oxygen acceptor. Annals Clin Biochem. 1969; 6(1): 24-27. https://doi.org/10.1177/000456326900600108
- Friedewald WT, Levy RI, Fredrickson DS. Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. Clin Chem. 1972; 18(6): 499-502. <u>https://doi.org/10.1093/clinchem/18.6.499</u>
- Canli EG, Dogan A, Canli M. Serum biomarker levels alter following nanoparticle (Al₂O₃, CuO, TiO₂) exposures in freshwater fish (Oreochromis niloticus). Environ Toxicol Pharmacol. 2018; 62: 181-187. <u>https://doi.org/10.1016/j.etap.2018.07.009</u>
- 34. Qassim B, Heino M, Morabito D. Uptake of Three Pharmaceuticals by Beans (Phaseolus vulgaris L.) from Contaminated Soils. Baghdad Sci J. 2020; 17(3): 0733-0742. http://dv.doi.org/10.21122/bri.2020.17.2.0723

http://dx.doi.org/10.21123/bsj.2020.17.3.0733

- 35. Fuentes-Gandara F, Pinedo-Hernández J, Marrugo-Negrete J, Díez S. Human health impacts of exposure to metals through extreme consumption of fish from the Colombian Caribbean Sea. Environ Geochem Health. 2018; 40(1): 229-242. https://doi.org/10.1007/s10653-016-9896-z
- 36. Pereira BMP, Wang X, Tagkopoulos I. Biocideinduced emergence of antibiotic resistance in Escherichia coli. Front Microbiol. 2021; 12: 640923. <u>https://doi.org/10.3389/fmicb.2021.640923</u>
- 37. Kampf G. Challenging biocide tolerance with antiseptic stewardship. J Hosp Infect. 2018 Sep 11; 100(3): e37-e39. http://doi.org/10.1016/j.jhin.2018.07.014
- 38. Deutschle T, Porkert U, Reiter R, Keck T, Riechelmann H. In vitro genotoxicity and cytotoxicity of benzalkonium chloride. Toxicol In

2023, 20(3 Suppl.): 945-956 https://dx.doi.org/10.21123/bsj.2023.7961 P-ISSN: 2078-8665 - E-ISSN: 2411-7986

Vitro. 2006; 20(8): 1472-1477. http://doi.org/10.1016/j.tiv.2006.07.006

- Omojowo FS, Sogbesan OA. Fish losses due to bacterial flora and infections of fishes in Kainji Lake Area, Nigeria: A Review. Niger Vet J. 2005; 24(2): 41-47. <u>http://doi.org/10.4314/nvj.v24i2.3453</u>
- 40. Mahboub HH, Khedr MH, Elshopakey GE, Shakweer MS, Mohamed DI, Ismail TA, et al. Impact of silver nanoparticles exposure on neurobehavior, hematology, and oxidative stress biomarkers of African catfish (Clarias gariepinus). Aquaculture 2021; 544: 737082. https://doi.org/10.1016/j.aquaculture.2021.737082
- 41. Gallego-Ríos SE, Peñuela GA, Martínez-López E. Updating the use of biochemical biomarkers in fish for the evaluation of alterations produced by pharmaceutical products. Environ Toxicol Pharmacol. 2021; 88: 103756. https://doi.org/10.1016/j.etap.2021.103756
- 42. Ahmed I, Reshi QM, Fazio F. The influence of the endogenous and exogenous factors on hematological parameters in different fish species: a review. Aquac internat. 2020; 28(3): 869-899. https://doi.org/10.1007/s10499-019-00501-3
- 43. Hertika AMS, Supriatna S, Darmawan A, Nugroho BA, Handoko AD, Qurniawatri AY, et al. The hematological profile of Barbonymus altus to evaluate water quality in the Badher bank conservation area, Blitar, East Java, Indonesia. Biodiversitas. 2021; 22(5): 2532-2541. https://doi.org/10.13057/biodiv/d220510
- 44. Hemalatha D, Muthukumar A, Rangasamy B, Nataraj B, Ramesh M. Impact of sublethal concentration of a fungicide propiconazole on certain health biomarkers of Indian major carp Labeo rohita. *Biocatal Agric Biotechnol*. 2016; 8: 321-7. <u>https://doi.org/10.1016/j.bcab.2016.10.009</u>
- 45. Ucar A, Özgeriş FB, Yeltekin AÇ, Parlak V, Alak G, Keleş MS, et al. The effect of N-acetylcysteine supplementation on the oxidative stress levels, apoptosis, DNA damage, and hematopoietic effect in pesticide-exposed fish blood. *J Biochem Mol Toxicol.* 2019; 33(6): e22311. https://doi.org/10.1002/jbt.22311
- 46. Said REM, Ashry M, AbdAllah EM. The use of biomarkers in the Nile Tilapia (Oreochromis niloticus) as biological signals to track Nile contamination in Egypt. Egypt J Aquat Biol Fish. 2021; 25(5): 203-214. http://doi.org/10.21608/ejabf.2021.198551
- 47. Soliman HAM, Hamed M, Sayed AEH. Investigating the effects of copper sulfate and copper oxide nanoparticles in Nile tilapia (Oreochromis niloticus) using multiple biomarkers: the

prophylactic role of Spirulina. Environ Sci Pollut Res Int. 2021; 28(23): 30046-30057. http://doi.org/10.1007/s11356-021-12859-0

- Hamed M, Osman AGM, Badrey AEA, Soliman HAM, Sayed AEH. Microplastics-Induced Eryptosis and Poikilocytosis in Early-Juvenile Nile Tilapia (Oreochromis niloticus). Front Physiol. 2021; 12: 742922. <u>http://doi.org/10.3389/fphys.2021.742922</u>
- Burgos-Aceves MA, Lionetti L, Faggio C. Multidisciplinary haematology as prognostic device in environmental and xenobiotic stress-induced response in fish. Sci Total Environ. 2019; 670: 1170-1183.

https://doi.org/10.1016/j.scitotenv.2019.03.275

- 50. Shah N, Khisroon M, Shah SSA. Assessment of copper, chromium, and lead toxicity in fish (Ctenopharyngodon idella Valenciennes, 1844) through hematological biomarkers. Environ Sci Pollut Res. 2020; 27(26): 33259-33269. https://doi.org/10.1007/s11356-020-09598-z
- 51. Gouda AMR, Hagras AE, Okbah MA, El-Gammal MI. Influence of the Linear Alkylbenzene Sulfonate (LAS) on hematological and biochemical parameters of Nile Tilapia, Oreochromis niloticus. Saudi J Biol Sci. 2022; 29(2): 1006-1013. http://doi.org/10.1016/j.sjbs.2021.09.074
- 52. Misra V, Kumar V, Pandey SD, Viswanathan P. Carbohydrate metabolism changes in fish fingerlings and yearlings exposed to linear alkyl benzene sulphonate. Water Air Soil Pollut. 1990; 50(3): 233-239. https://doi.org/10.1007/bf00280625
- 53. Feng R, Ma LJ, Wang M, Liu C, Yang R, Su H, et al. Oxidation of fish oil exacerbates alcoholic liver disease by enhancing intestinal dysbiosis in mice. Commun Biol. 2020; 3: 481. https://doi.org/10.1038/s42003-020-01213-8
- 54. Monfared AL, Salati AP. Histomorphometric and biochemical studies on the liver of rainbow trout (Oncorhynchus mykiss) after exposure to sublethal concentrations of phenol. Toxicol Ind Health. 2013; 29(9): 856-861.

https://doi.org/10.1177/0748233712451765

55. Osman AGM, AbouelFadl KY, Abd El Reheem AEBM, Mahmoud UM, Kloas W, Moustafa MA. Blood Biomarkers in Nile tilapia Oreochromis niloticus niloticus and African Catfish Clarias gariepinus to Evaluate Water Quality of the River Nile. J fish sci com 2018; 12(1). http://doi.org/10.21767/1307-234x.1000141



- 56. Herron JM, Hines KM, Tomita H, Seguin RP, Cui JY, Xu L. Multi-omics investigation reveals benzalkonium chloride disinfectants alter sterol and lipid homeostasis in the mouse neonatal brain. Toxicol Sci. 2019; 171(1): 32-45. http://doi.org/10.1093/toxsci/kfz139
- 57. Gabriel U, George A. Plasma enzymes in Clarias gariepinus exposed to chronic levels of round up (glyphosate). Environ Ecol.2005; 23(2): 271-276. https://ssrn.com/abstract=3201447
- 58. Lee S, Saravanan M, Kim S-A, Rhee J-S. Long-term exposure to antifouling biocide chlorothalonil

modulates immunity and biochemical and antioxidant parameters in the blood of olive flounder. Comp. Biochem. Physiol. C,: Toxicol Pharmacol. 2022; 257: 109337. https://doi.org/10.1016/j.cbpc.2022.109337

59. Abbas H, El-Badawi A. Use of hematological and biochemical parameters and histological changes to assess the toxicity of drumstick tree (Moringa oleifera) seeds extract on Tilapia (Oreochromis niloticus) fish. Egypt J Aquat Biol Fish. 2014; 18(3): 21-40. https://doi.org/10.21608/ejabf.2014.2215

النتائج المصلية الدموية كإشارات مبكرة في البلطي النيلي اوريكروميس نيلوتيكس المعالج بكلوريد البنز الكونيوم

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الخلاصة

الهدف الرئيسي من هذه الدراسة هو تقييم سمية كلوريد البنز الكونيوم في الاستزراع المائي ، باستخدام المؤشرات المصلية الدموية لسمكة البلطي النيلي Oreochromis niloticus كمؤشرات حيوية. بعدما تعرضت الأسماك لثلاث مستويات من كلوريد البنز الكونيوم 0.1 ، 0.25 ، 0.50 ، و 1 ملغم / لتر في الأحواض المائية لفترتين زمنيتين 21 و 42 يومًا ، تم تقييم الفحص الميكروبيولوجي في أحواض الأسماك ، بالإضافة إلى معايير الدم. أشارت نتائج الفحص البكتيري أنه باستثناء متوسط الفرق بين المعاملة الثانية والثالثة من كلوريد البنزالكونيوم (فرق غير معنوي – 42 يوم) فقد أظهرت النتائج فروقاً كبيرة عند مقارنة باقى المتوسطات في جميع المعاملات ($0.05 \ge 0.01$). انخفضت مؤشرات الدم الرئيسية ككرات الدم الحمراء والهيموجلوبين بشكل متفاوت عقب التعرض للثلاث تركيزات الأولي عند مقارنتها بمجموعة السيطرة، ثم بلغت ذروتها عند أعلي تركيز من كلوريد البنز الكونيوم رغم عدم وجود فروق احصائية عند مقارنتها بمجموعة السيطرة (باستثناء الهيموجلوبين – 42 يوم) . على العكس من ذلك ، ارتفعت كرات الدم البيضاء بشكل سريع عند التركيز الأول من كلوريد البنز الكونيوم خاصة في الفترة 42 يوما بالمقارنةً نتائج مجموعة السيطرة. تغيرت متوسطات كرات الدم البيضاء بعد التعرض للتركيزين الثاني و الثالث ، قبل أن تبلغ ذروتها عند أعلى تركيز من كلوريد البنز الكونيوم. أظهرت MCV و MCH و MCHC تقلبًا طغيفًا بين مجموعة السيطرة والأسماك المعالجة . فيما يتعلق بالنتائج البيوكيميائية ، أظهر متوسط مستويات الكوليسترول والدهون الثلاثية نمطأ غير منتظم حسب تركيز المعالجة الكيميائية. تقلبت مستويات ALT و AST بين أسماك مجموعة السيطرة و مجموعة المعالجة الكيميانية الثلاث الأولى ، قبل أن تزداد عند التركيز الأعلى مع وجود فروق معنوية(باستثناءAST عند مقارنة التركيزين 3 و4 من كلوريد البنزالكونيوم عقب 42 يوم حيث كانت الفروق للمتوسطات غير معنوية من الناحية الاحصائية) عند مقارنة مجموعة السيطرة , BAC1,2,3 بالمجموعة التي تعرضت ل BAC4. بعد التعرض لكلوريد البنز الكونيوم ، بقيت المعلمات الأخرى ، بما في ذلك البروتين واليوريا كما هي في مجموعة السيطرة أشار التغيير في بعض علامات الدم المصلية على أن هناك استجابة حيوية فيسيولوجية للإجهاد الناجم عن التعرض للمبيدات الحيوية.

الكلمات المفتاحية: كلوريد البنز الكونيوم ، المؤشر ات الحيوية ، المطهر ، Oreochromis niloticus ، السمية.